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UNITED STATES PATENT APPLICATION

OF

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FOR

HUMAN BETA-1,3-GALACTOSYLTRANSFERASE

Description

BETA-1,3-GALACTOSYLTRANSFERASE HOMOLOG, ZNSSP6

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REFERENCE TO RELATED APPLICATIONS

This application is related to Provisional Application 60/115,721 filed on January 12, 1999. Under 35 U.S.C. § 119(e)(1), this application claims benefit of said Provisional Application.

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BACKGROUND OF THE INVENTION

Glycosyltransferase molecules transfer carbohydrate molecules to glycoproteins during biosynthesis. Members of this family have also been detected on the cell surface where they are thought to be involved in varying aspects of cell-cell interactions. This family includes carbohydrate transferring enzymes, such as sialyltransferases and fucosyltransferases, and galactosyltransferases. During the formation of *O*-linked glycoproteins and the modification of *N*-linked ones, each sugar transfer is catalyzed by a different type of glycosyltransferase. Each glycosyltransferase enzyme is specific for both the donor sugar nucleotide and the acceptor molecule.

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Galactosyltransferases promote the transfer of an activated galactose residue in UDP-galactose to the monosaccharide *N*-acetylglucosamine. This transfer is a step in the biosynthesis of the carbohydrate portion of galactose-containing glycoproteins, such as oligosaccharides and glycolipids, in animal tissues. One subgroup of the galactosyltransferases is the beta-1,3-galactosyltransferases, which are characterized by the elongation of type I oligosaccharide chains. Additionally, the beta-1,3-galactosyltransferases are found on glycoproteins and glycolipids, are important precursors of blood group antigens, and are present in soluble oligosaccharides of human milk. Similar to other members of galactosyltransferases. The beta-1,3-galactosyltransferases require a divalent cation (Mn^{2+}) to function. The beta-1,3-galactosyltransferases seem to have restricted tissue distributions.

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Some galactosyltransferases are found in the Golgi apparatus. These Golgi-localized enzymes have structure similarity: a short N-terminal domain that faces the cytosol, a single transmembrane α helix, and a large C-terminal domain that faces the Golgi lumen and that contains the catalytic site. The transmembrane α helix is necessary and sufficient to restrict the enzyme to the Golgi. Of the beta-1,3-galactosyltransferase family two members (See Amado, M. et al., J. Biol. Chem. 273, 21: 12770-12778, 1998) have been predicted to have two potentially different initiation codons, resulting in two different N-terminal cytoplasmic domains.

Additionally, galactosyltransferases have been shown to be expressed on the cell surface, where their function is theorized to participate in cellular interactions, perhaps as receptors, or receptor-like complementary molecules as well as secreted ligands. As a cell surface carbohydrate, galactosyltransferases have been implicated in varied biology such as cell migration, contact inhibition, tissue interactions, neuronal specificity, fertilization, embryonic cell adhesions, limb bud morphogenesis, mesenchyme development, immune recognition, growth control, and tumor metastasis. See, for example, Shur, B.D., Mol Cell Bioc. 61:143-158, 1984.

The failure of tumor cell-tumor cell adhesion is believed to be a contributing factor in tumor metastases. See, for example, Zetter, Cancer Biology, 4: 219-29, 1993. Metastases, in turn, are generally associated with poor prognosis for cancer treatment. The metastatic process involves a variety of cellular events, including angiogenesis, tumor cell invasion of the vascular or lymphatic circulation, tumor cell arrest at a secondary site; tumor cell passage across the vessel wall into the parenchymal tissue, and tumor cell proliferation at the secondary site. Thus, both positive and negative regulation of adhesion are necessary for metastasis. That is, tumor cells must break away from the primary tumor mass, travel in circulation and adhere to cellular and/or extracellular matrix elements at a secondary site. Molecules capable of modulating cell-cell and cell-matrix adhesion are therefore sought for the study, diagnosis, prevention and/or treatment of metastases.

Beta-1,3-galactosyltransferases have limited homology to each other. In contrast to other glycosyltransferases, they do not appear to be localized to the same chromosomes. Additionally, a member of this family has recently been identified in

Drosophila. This molecule, *Brainiac* (*brn*), also known as a Neurogenic Secreted Signaling Peptide (NSSP), is involved in contact and adhesion between germ-line and follicle cells (Amado, M. et al., J. Biol. Chem. 273, 21: 12770-12778, 1998). Germline *Brainiac* activity has been shown to be essential for development of follicular epithelium (Goode, S. et al., Dev. Biol. 178:35-50, 1996). Additionally, *brn* is required continuously throughout oogenesis, beginning in the germarium at the time that follicle cells envelop the oocyte-nurse cell complex and continuing stages when the eggshell is produced. The expression of *brn* in the germline continuously throughout oogenesis is consistent with *brn*'s role in developing the follicular epithelium around each germline cyst, as well as for dorsal-ventral patterning of the follicular epithelium during later phases of oogenesis. See Goode, S. et al., Development. 116: 177-192, 1992.

A deficiency of beta-1,3-galactosyltransferase enzymes has been noticed in the Tn-syndrome. This syndrome is a rarely acquired disorder affecting all hemopoietic lineages, and is characterized by the expression of the Tn and the sialosyl-Tn antigens on the cell surface. The Tn is α N-acetylgalactosamine linked O-glycosidically to threonine or serine residues of membrane proteins. These antigens bind naturally occurring serum antibodies thereby leading to mild hemolytic anemia and pronounced thrombopenia. Thus, the blood cells in the Tn-syndrome are expected to carry less sialic acid if galactose can not be transferred to N-Acetylgalactosamine. The expression of Tn and sialosyl-Tn antigens as a consequence of incomplete or disordered glycan biosynthesis has been recognized as a cancer-associated phenomenon. Tn and sialosyl-Tn antigens are among the most investigated cancer-associated carbohydrate antigens.

The present invention provides such polypeptides for these and other uses that should be apparent to those skilled in the art from the teachings herein.

SUMMARY OF THE INVENTION

Within one aspect, the invention provides an isolated polypeptide comprising residues 114 to 370 of SEQ ID NO:2. Within an embodiment, the isolated polypeptide comprises residues 114 to 378 of SEQ ID NO:2. Within another embodiment, the isolated polypeptide comprises residues 50 to 378 of SEQ ID NO:2. Within another embodiment, the isolated polypeptide comprises residues 26 to 378 of

SEQ ID NO:2. Within another embodiment, the isolated polypeptide comprises residues 1 to 378 of SEQ ID NO:2.

Within another aspect, the invention provides an isolated polypeptide selected from the group consisting of: a) a polypeptide comprising residues 1 to 25 of
5 SEQ ID NO:2; b) a polypeptide comprising residues 26 to 49 of SEQ ID NO:2; c) a polypeptide comprising residues 50 to 113 of SEQ ID NO:2; d) a polypeptide comprising residues 114 to 370 of SEQ ID NO:2; e) a polypeptide comprising residues 371 to 378 of SEQ ID NO:2; and f) a polypeptide comprising residues 1 to 378 of SEQ ID NO:2.

10 Within another aspect, the invention provides an isolated polynucleotide encoding a polypeptide wherein the polypeptide comprises residues 114 to 370 of SEQ ID NO:2. Within another embodiment, the isolated polynucleotide comprises residues 114 to 378 of SEQ ID NO:2. Within another embodiment, the isolated polynucleotide comprises residues 50 to 378 of SEQ ID NO:2. Within another embodiment, the
15 isolated polynucleotide comprises residues 26 to 378 of SEQ ID NO:2. Within another embodiment, the isolated comprises residues 1 to 378 of SEQ ID NO:2.

Within another aspect, the invention provides an isolated polynucleotide encoding a polypeptide molecule wherein the polypeptide is selected from the group consisting of: a) a polypeptide comprising residues 1 to 25 of SEQ ID NO:2; b) a
20 polypeptide comprising residues 26 to 49 of SEQ ID NO:2; c) a polypeptide comprising residues 50 to 113 of SEQ ID NO:2; d) a polypeptide comprising residues 114 to 370 of SEQ ID NO:2; e) a polypeptide comprising residues 371 to 378 of SEQ ID NO:2; and f) a polypeptide comprising residues 1 to 378 of SEQ ID NO:2.

Within another aspect, the invention provides an expression vector
25 comprising the following operably linked elements: a transcription promoter; a DNA segment wherein the DNA segment is a polynucleotide encoding the polypeptide of claim 1; and a transcription terminator. Within another embodiment, the DNA segment contains an affinity tag. Within another embodiment, the invention provides a cultured cell into which has been introduced the expression vector wherein said cell expresses
30 the polypeptide encoded by the DNA segment. Within another embodiment, the invention provides a method of producing a polypeptide comprising culturing a cell,

whereby said cell expresses the polypeptide encoded by the DNA segment; and recovering the polypeptide. Within another embodiment, the invention provides the polypeptide produced by said method of expression.

Within another aspect, the invention provides a method of producing an antibody comprising the following steps in order: inoculating an animal with an epitope bearing portion of a polypeptide wherein the epitope bearing portion is selected from the group consisting of: a polypeptide comprising residues 10 to 16 of SEQ ID NO:2; a polypeptide comprising residues 52 to 61 of SEQ ID NO:2; a polypeptide comprising residues 52 to 78 of SEQ ID NO:2; a polypeptide comprising residues 69 to 78 of SEQ ID NO:2; a polypeptide comprising residues 89 to 94 of SEQ ID NO:2; a polypeptide comprising residues 89 to 117 of SEQ ID NO:2; a polypeptide comprising residues 111 to 117 of SEQ ID NO:2; a polypeptide comprising residues 126 to 134 of SEQ ID NO:2; a polypeptide comprising residues 126 to 151 of SEQ ID NO:2; a polypeptide comprising residues 143 to 151 of SEQ ID NO:2; a polypeptide comprising residues 215 to 220 of SEQ ID NO:2; a polypeptide comprising residues 215 to 239 of SEQ ID NO:2; a polypeptide comprising residues 223 to 239 of SEQ ID NO:2; a polypeptide comprising residues 223 to 257 of SEQ ID NO:2; a polypeptide comprising residues 251 to 257 of SEQ ID NO:2; and a polypeptide comprising residues 332 to 337 of SEQ ID NO:2; wherein the polypeptide elicits an immune response in the animal to produce the antibody; and isolating the antibody from the animal. Within an embodiment, the antibody produced binds to a polypeptide comprising residues 114 to 370 of SEQ ID NO:2. Within another embodiment, the antibody is a monoclonal antibody. Within another embodiment, the antibody is joined to a moiety selected from the group consisting of: a) an affinity tag; b) a detectable molecule; c) a cytotoxic molecule; and d) a cytokine. Within another embodiment, the invention provides a method of killing cells expressing a polypeptide comprising residues 114 to 370 of SEQ ID NO:2, comprising contacting the cells with the antibody joined to the moiety, wherein the antibody is joined to the cytotoxic molecule.

Within another aspect, the invention provides a method for modulating cell-cell interactions by combining a polypeptide comprising residues 114 to 370 of SEQ ID NO:2 with cells. Within another embodiment, the cells are derived from

tissues selected from the group consisting of: a) tissues from brain; b) issues from kidney; and c) issues from testis.

Within another aspect, the invention provides a method for modulating glycoprotein and glycolipid biosynthesis in cells, cell culture, and cell matrix comprising contacting the cells, cell culture or cell matrix with the polypeptide comprising residues 114 to 370 of SEQ ID NO:2. Within an embodiment, the cells are derived from tissues selected from the group consisting of: a) tissues from brain; b) issues from kidney; and c) tissues from testis.

Within another aspect, the invention provides a method of detecting a znssp6 anti-complementary molecule comprising contacting a test sample containing the znssp6 anti-complementary molecule with a polypeptide comprises residues 114 to 370 of SEQ ID NO:2.

Within another aspect, the present invention provides an isolated polynucleotide molecule encoding a polypeptide wherein said polynucleotide molecule is selected from the group consisting of: a) polynucleotide molecules comprising a nucleotide sequence from nucleotide 474 to nucleotide 1244 of SEQ ID NO: 1; b) polynucleotide molecules that encode a polypeptide comprising a sequence of amino acid residues that is at least 70% identical to amino acid residues 114 to 370 of SEQ ID NO: 2; c) degenerate nucleotide sequences of a), or b); and d) polynucleotide molecules that have a complementary sequence to a), b), or c). Within an embodiment, the polypeptide differs from amino acid residues 114 to 370 of SEQ ID NO:2 by conservative amino acid substitutions. Within another embodiment, the polypeptide consists of the sequence of SEQ ID NO:2. Within a further embodiment, the polypeptide is the sequence of amino acid residues 114 to 370 of SEQ ID NO:2. Within a related embodiment, is provided an isolated antibody or antibody fragment that specifically binds to the polypeptide.

Within another aspect, the present invention provides an isolated polynucleotide molecule encoding a polypeptide wherein said polynucleotide molecule is selected from the group consisting of: a) polynucleotide molecules comprising a nucleotide sequence from nucleotide 282 to nucleotide 1244 of SEQ ID NO: 1; b) polynucleotide molecules that encode a polypeptide comprising a sequence of amino

acid residues that is at least 70% identical to amino acid residues 50 to 370 of SEQ ID NO: 2; c) degenerate nucleotide sequences of a), or b); and d) polynucleotide molecules that have a complementary sequence to a), b), or c). Within an embodiment, the polypeptide differs from amino acid residues 50 to 370 of SEQ ID NO:2 by
5 conservative amino acid substitutions.

Within another aspect, the present invention provides an isolated polynucleotide molecule encoding a polypeptide wherein said polynucleotide molecule is selected from the group consisting of: a) polynucleotide molecules comprising a nucleotide sequence from nucleotide 135 to nucleotide 1268 of SEQ ID NO: 1; b)
10 polynucleotide molecules that encode a polypeptide comprising a sequence of amino acid residues that is at least 70% identical to amino acid residues 1 to 378 of SEQ ID NO: 2; c) degenerate nucleotide sequences of a), or b); and d) polynucleotide molecules that have a complementary sequence to a), b), or c). Within an embodiment, the polypeptide differs from amino acid residues 1 to 370 SEQ ID NO:2 by conservative
15 amino acid substitutions.

Within another aspect, the present invention provides, an expression vector comprising the following operably linked elements: a transcription promoter; a DNA segment encoding a polypeptide, wherein the DNA segment is selected from the group consisting of: a) polynucleotide molecules comprising a nucleotide sequence
20 from nucleotide 474 to nucleotide 1244 of SEQ ID NO: 1; b) polynucleotide molecules that encode a polypeptide that is at least 70% identical to amino acid residues 114 to 370 of SEQ ID NO: 2; and c) degenerate nucleotide sequences of a), or b); and a transcription terminator. Within an embodiment, the polypeptide differs from amino acid residues 114 to 370 SEQ ID NO:2 by conservative amino acid substitutions. Also
25 is provided a cultured cell into which has been introduced an expression vector, as described above, wherein said cell expresses the polypeptide encoded by the DNA segment.

Within another aspect, the present invention provides a method of producing a polypeptide comprising culturing a cell, whereby said cell expresses the
30 polypeptide encoded by the DNA segment as described above; and recovering the polypeptide.

Within another aspect, the present invention provides a method for modulating cell-cell interactions by combining a polypeptide, that is at least 70% identical to amino acid residues 114 to 370 SEQ ID NO:2, with cells *in vitro* and *in vivo*.

5 Within another aspect, the present invention provides a pharmaceutical composition comprising a polypeptide, that is at least 70% identical to amino acid residues 114 to 370 SEQ ID NO:2, in combination with a pharmaceutically acceptable vehicle.

10 Within another aspect, the present invention provides an isolated polynucleotide molecule encoding a fusion protein comprising a beta-1,3-galactosyltransferase domain having the amino acid sequence of residues 114 to 370 of SEQ ID NO:2, wherein said beta-1,3-galactosyltransferase domain is operably linked to an additional polypeptide. Within an embodiment, the additional polypeptide is selected from the group consisting of: a) affinity tag polypeptide molecules; b) 15 immunoglobulin heavy chain constant region polypeptide molecules; c) the hydrophobic region of amino acid residues 26 to 49 of SEQ ID NO:2; and d) hydrophobic regions of other beta-1,3-galactosyltransferase polypeptide molecules.

DETAILED DESCRIPTION OF THE INVENTION

20 Prior to setting forth the invention in detail, it may be helpful to the understanding thereof to define the following terms:

25 The term "affinity tag" is used herein to denote a polypeptide segment that can be attached to a second polypeptide to provide for purification of the second polypeptide or provide sites for attachment of the second polypeptide to a substrate. In principal, any peptide or protein for which an antibody or other specific binding agent is available can be used as an affinity tag. Affinity tags include a poly-histidine tract, protein A (Nilsson et al., EMBO J. 4:1075, 1985; Nilsson et al., Methods Enzymol. 198:3, 1991), glutathione S transferase (Smith and Johnson, Gene 67:31, 1988), Glu-Glu affinity tag (Grussenmeyer et al., Proc. Natl. Acad. Sci. USA 82:7952-4, 1985), substance P, Flag™ peptide (Hopp et al., Biotechnology 6:1204-1210, 1988), 30 streptavidin binding peptide, maltose binding protein (Guan et al., Gene 67:21-30, 1987), cellulose binding protein, thioredoxin, ubiquitin, T7 polymerase, or other

antigenic epitope or binding domain. See, in general, Ford et al., Protein Expression and Purification 2: 95-107, 1991. DNAs encoding affinity tags and other reagents are available from commercial suppliers (e.g., Pharmacia Biotech, Piscataway, NJ; New England Biolabs, Beverly, MA; Eastman Kodak, New Haven, CT).

5 The term "allelic variant" is used herein to denote any of two or more alternative forms of a gene occupying the same chromosomal locus. Allelic variation arises naturally through mutation, and may result in phenotypic polymorphism within populations. Gene mutations can be silent (no change in the encoded polypeptide) or may encode polypeptides having altered amino acid sequence. The term allelic variant
10 is also used herein to denote a protein encoded by an allelic variant of a gene.

 The terms "amino-terminal" and "carboxyl-terminal" are used herein to denote positions within polypeptides. Where the context allows, these terms are used with reference to a particular sequence or portion of a polypeptide to denote proximity or relative position. For example, a certain sequence positioned carboxyl-terminal to a
15 reference sequence within a polypeptide is located proximal to the carboxyl terminus of the reference sequence, but is not necessarily at the carboxyl terminus of the complete polypeptide.

 The term "complement/anti-complement pair" denotes non-identical moieties that form a non-covalently associated, stable pair under appropriate conditions.
20 For instance, biotin and avidin (or streptavidin) are prototypical members of a complement/anti-complement pair. Other exemplary complement/anti-complement pairs include receptor/ligand pairs, antibody/antigen (or hapten or epitope) pairs, sense/antisense polynucleotide pairs, and the like. Where subsequent dissociation of the complement/anti-complement pair is desirable, the complement/anti-complement
25 pair preferably has a binding affinity of $<10^9 \text{ M}^{-1}$.

 The term "complements of a polynucleotide molecule" is a polynucleotide molecule having a complementary base sequence and reverse orientation as compared to a reference sequence. For example, the sequence 5' ATGCACGGG 3' is complementary to 5' CCCGTGCAT 3'.

30 The term "contig" denotes a polynucleotide that has a contiguous stretch of identical or complementary sequence to another polynucleotide. Contiguous

sequences are said to "overlap" a given stretch of polynucleotide sequence either in their entirety or along a partial stretch of the polynucleotide. For example, representative contigs to the polynucleotide sequence 5'-ATGGAGCTT-3' are 5'-AGCTTgagt-3' and 3'-tcgacTACC-5'.

5 The term "degenerate nucleotide sequence" denotes a sequence of nucleotides that includes one or more degenerate codons (as compared to a reference polynucleotide molecule that encodes a polypeptide). Degenerate codons contain different triplets of nucleotides, but encode the same amino acid residue (i.e., GAU and GAC triplets each encode Asp).

10 The term "expression vector" is used to denote a DNA molecule, linear or circular, that comprises a segment encoding a polypeptide of interest operably linked to additional segments that provide for its transcription. Such additional segments include promoter and terminator sequences, and may also include one or more origins of replication, one or more selectable markers, an enhancer, a polyadenylation signal,
15 etc. Expression vectors are generally derived from plasmid or viral DNA, or may contain elements of both.

 The term "isolated", when applied to a polynucleotide, denotes that the polynucleotide has been removed from its natural genetic milieu and is thus free of other extraneous or unwanted coding sequences, and is in a form suitable for use within
20 genetically engineered protein production systems. Such isolated molecules are those that are separated from their natural environment and include cDNA and genomic clones. Isolated DNA molecules of the present invention are free of other genes with which they are ordinarily associated, but may include naturally occurring 5' and 3' untranslated regions such as promoters and terminators. The identification of
25 associated regions will be evident to one of ordinary skill in the art (see for example, Dynan and Tijan, Nature 316:774-78, 1985).

 An "isolated" polypeptide or protein is a polypeptide or protein that is found in a condition other than its native environment, such as apart from blood and animal tissue. In a preferred form, the isolated polypeptide is substantially free of other
30 polypeptides, particularly other polypeptides of animal origin. It is preferred to provide the polypeptides in a highly purified form, i.e. greater than 95% pure, more preferably

greater than 99% pure. When used in this context, the term "isolated" does not exclude the presence of the same polypeptide in alternative physical forms, such as dimers or alternatively glycosylated or derivatized forms.

The term "joined" when referring to attaching one moiety to another moiety, indicates that the moieties are directly or indirectly linked to each other. Thus, such moieties can be joined by genetic manipulations (such as, for example, fusion proteins), chemical manipulations (such as, for example, chemical conjugation, coupling, or chelation), or other means.

The term "operably linked", when referring to DNA segments, indicates that the segments are arranged so that they function in concert for their intended purposes, e.g., transcription initiates in the promoter and proceeds through the coding segment to the terminator.

The term "ortholog" or "species homolog", denotes a polypeptide or protein obtained from one species that is the functional counterpart of a polypeptide or protein from a different species. Sequence differences among orthologs are the result of speciation.

"Paralogs" are distinct but structurally related proteins made by an organism. Paralogs are believed to arise through gene duplication. For example, α -globin, β -globin, and myoglobin are paralogs of each other.

A "polynucleotide" is a single- or double-stranded polymer of deoxyribonucleotide or ribonucleotide bases read from the 5' to the 3' end. Polynucleotides include RNA and DNA, and may be isolated from natural sources, synthesized *in vitro*, or prepared from a combination of natural and synthetic molecules. Sizes of polynucleotides are expressed as base pairs (abbreviated "bp"), nucleotides ("nt"), or kilobases ("kb"). Where the context allows, the latter two terms may describe polynucleotides that are single-stranded or double-stranded. When the term is applied to double-stranded molecules it is used to denote overall length and will be understood to be equivalent to the term "base pairs". It will be recognized by those skilled in the art that the two strands of a double-stranded polynucleotide may differ slightly in length and that the ends thereof may be staggered as a result of enzymatic cleavage; thus all

nucleotides within a double-stranded polynucleotide molecule may not be paired. Such unpaired ends will in general not exceed 20 nt in length.

A "polypeptide" is a polymer of amino acid residues joined by peptide bonds, whether produced naturally or synthetically. Polypeptides of less than about 10 amino acid residues are commonly referred to as "peptides".

The term "promoter" is used herein for its art-recognized meaning to denote a portion of a gene containing DNA sequences that provide for the binding of RNA polymerase and initiation of transcription. Promoter sequences are commonly, but not always, found in the 5' non-coding regions of genes.

A "protein" is a macromolecule comprising one or more polypeptide chains. A protein may also comprise non-peptidic components, such as carbohydrate groups. Carbohydrates and other non-peptidic substituents may be added to a protein by the cell in which the protein is produced, and will vary with the type of cell. Proteins are defined herein in terms of their amino acid backbone structures; substituents such as carbohydrate groups are generally not specified, but may be present nonetheless.

The term "receptor" denotes a cell-associated protein that binds to a bioactive molecule (i.e., a ligand) and mediates the effect of the ligand on the cell. Membrane-bound receptors are characterized by a multi-domain or multi-peptide structure comprising an extracellular ligand-binding domain and an intracellular effector domain that is typically involved in signal transduction. Binding of ligand to receptor results in a conformational change in the receptor that causes an interaction between the effector domain and other molecule(s) in the cell. This interaction in turn leads to an alteration in the metabolism of the cell. Metabolic events that are linked to receptor-ligand interactions include gene transcription, phosphorylation, dephosphorylation, increases in cyclic AMP production, mobilization of cellular calcium, mobilization of membrane lipids, cell adhesion, hydrolysis of inositol lipids and hydrolysis of phospholipids. In general, receptors can be membrane bound, cytosolic or nuclear; monomeric (e.g., thyroid stimulating hormone receptor, beta-adrenergic receptor) or multimeric (e.g., PDGF receptor, growth hormone receptor, IL-

3 receptor, GM-CSF receptor, G-CSF receptor, erythropoietin receptor and IL-6 receptor).

The term "secretory signal sequence" denotes a DNA sequence that encodes a polypeptide (a "secretory peptide") that, as a component of a larger polypeptide, directs the larger polypeptide through a secretory pathway of a cell in which it is synthesized. The larger polypeptide is commonly cleaved to remove the secretory peptide during transit through the secretory pathway.

A "segment" is a portion of a larger molecule (e.g., polynucleotide or polypeptide) having specified attributes. For example, a DNA segment encoding a specified polypeptide is a portion of a longer DNA molecule, such as a plasmid or plasmid fragment, that, when read from the 5' to the 3' direction, encodes the sequence of amino acids of the specified polypeptide.

The term "splice variant" is used herein to denote alternative forms of RNA transcribed from a gene. Splice variation arises naturally through use of alternative splicing sites within a transcribed RNA molecule, or less commonly between separately transcribed RNA molecules, and may result in several mRNAs transcribed from the same gene. Splice variants may encode polypeptides having altered amino acid sequence. The term splice variant is also used herein to denote a protein encoded by a splice variant of an mRNA transcribed from a gene.

Molecular weights and lengths of polymers determined by imprecise analytical methods (e.g., gel electrophoresis) will be understood to be approximate values. When such a value is expressed as "about" X or "approximately" X, the stated value of X will be understood to be accurate to $\pm 10\%$.

All references cited herein are incorporated by reference in their entirety.

The present invention is based upon the discovery of a novel cDNA sequence (SEQ ID NO:1) and corresponding polypeptide (SEQ ID NO:2) having homology to a family of proteins, the beta-1,3-galactosyltransferases. The beta-1,3-galactosyltransferases are part of the galactosyltransferases, which in turn, belong in the category of glycosyltransferases. The beta-1,3-galactosyltransferase family includes HSY15014 (Kolbinger, F. et al., Journal of Biol. Chem. 273: 433-440, 1998), HSGALT3, HSGALT4, (Amado, M. et al., *ibid*), E07739 (Katsutoshi, S. et al.,

Japanese patent, JP 1994181759-A/1), and Cardiac and Pancreatic Peptide (Human Genome Sciences, Inc., WO 98/44112). Enzymes in this category are responsible for transferring galactose to carbohydrate chains during biosynthesis. It has been predicted that the beta-1,3-galactosyltransferase family members are in the alpha/beta barrel (TIM barrel) folding class of enzymes, similar to other glycosyltransferases such as the alpha-amylases and beta-glycanases (Yuan, Y. et al., Cell 88:9-11, 1997). Another member of the beta-1,3-galactosyltransferase family is the *Drosophila melanogaster* locus Brainiac (*brn*) (Goode, S. et al., Devel. Biol. 178:35-50, 1996), also known as "putative neurogenic secreted signaling protein" or NSSP. *Brn* is required for epithelial development (Goode, *ibid*). This activity may be due to possible cell interactions between the membrane bound glycosyltransferase and oligosaccharide substrates on adjacent cell surfaces (Shur, *ibid*). The beta-1,3-galactosyltransferases family members are also known as neurogenic secreted signal peptides. See, for example, Shur, B.D., *ibid*, and Amado, M. et al., *ibid*.

The beta-1,3-galactosyltransferases are predicted to be Type II transmembrane proteins. An ortholog to E07739, is AF029790 (Hennet, T. et al., Journal of Biol. Chem. 273:58-65, 1998), which is claimed to be a Type II transmembrane domain based on hydrophobicity analysis. However, due to the close proximity of this domain to the initiating methionine and lack of positively charged residues preceding the domain it is possible that AF029790 is not membrane bound but rather an extracellular secreted protein.

The sequence of the novel znssp6 polypeptides of the present invention was initially identified by searching an EST database for open reading frames with similarity to *brn*. The insert of an expressed sequence tag was obtained and the deduced amino acid sequence of the insert was determined to be incomplete at the 5' end. Polymerase chain extensions of this sequence were performed, and their analysis identified a second EST for which the insert was obtained and sequenced. Analysis of the nucleotide sequence of the second insert (SEQ ID NO:1) revealed an open reading frame encoding 378 amino acids (SEQ ID NO: 2) which has been designated as znssp6.

A representative motif of the beta-1,3-galactosyltransferase family is described by the following amino acid residue profile: [D,E] [D] [V] [F,Y] [L,T,V]

[G]. The sequence of amino acid residues from residue 304 to 309 of SEQ ID NO:2 is representative of this motif. The core region of similarity to all beta-1,3-galactosyltransferases begins at residue 114 and ends at residue 370 of SEQ ID NO:2. This is believed to be the catalytic, or anti-complementary molecule binding domain.

5 Conserved negatively charged amino acid residues 174, 180, 185, 220, 304, and 305 of SEQ ID NO:2 are contained within this catalytic/binding domain. Amino acid residues 50 to 113 of SEQ ID NO:2 are predicted to form a stem or linker domain separating the catalytic/binding domain from the cell membrane. The region from residue 26 to residue 49 of SEQ ID NO:2 is strongly hydrophobic and in one form of the znssp6
10 protein, this region is predicted to form a transmembrane domain resulting in a membrane bound form of znssp6. In another form of the znssp6 protein, the hydrophobic domain may act as a secretory peptide in which case znssp6 is a secreted, soluble protein. Znssp6 shares homology with beta-1,3-galactosyltransferases which are predicted to be Type II membrane proteins. Znssp6 shows the highest similarity to
15 Cardiac and Pancreatic Peptide (CAPP, See WO 98/44112, 1998), at 43% amino acid identity. Additionally, znssp6 has 36% identity to *brn*. Those skilled in the art will recognize that predicted domain boundaries are approximations based on primary sequence content, and may vary slightly; however, such estimates are generally accurate to within ± 5 amino acid residues.

20 The present invention also provides post translationally modified polypeptides or polypeptide fragments. A conserved potential N-linked glycosylation site can be found at amino acid residue 192 of SEQ ID NO:2. Other potential N-linked glycosylation sites are at residues 79, and 104 of SEQ ID NO:2. Other examples of post translational modifications include proteolytic cleavage, disulfide bonding and
25 hydroxylation.

Analysis of the tissue distribution of znssp6 was performed by the Northern blotting technique using Human Multiple Tissue and Master Dot Blots. Strong signals were observed in brain (adult and fetal), kidney (adult and fetal), and testis. The major transcript size was about 1.6 kb, although minor transcripts of about
30 3.0 kb, and 4.0 kb were also evident. Lower level expression was also observed in tissues such as spinal cord, colon, prostate, stomach, ovary, pancreas, pituitary gland,

adrenal gland, salivary gland, mammary gland, liver, small intestine, spleen, thymus, peripheral leukocyte, lymph node, bone marrow, lung, trachea, placenta, fetal spleen and fetal lung.

The highly conserved, negatively charged residues at positions 174,
 5 180, 185, 220, 304, and 305 of SEQ ID NO:2 and the amino acid sequence between
 304 and 309 of znssp6 can be used to identify new family members. For instance,
 reverse transcription-polymerase chain reaction (RT-PCR) can be used to amplify
 sequences encoding the znssp6 polynucleotide from RNA obtained from a variety of
 tissue sources or cell lines. In particular, highly degenerate primers designed from the
 10 znssp6 sequences are useful for this purpose.

POLYNUCLEOTIDES:

The present invention also provides polynucleotide molecules, including
 DNA and RNA molecules, that encode the znssp6 polypeptides disclosed herein.
 Those skilled in the art will readily recognize that, in view of the degeneracy of the
 15 genetic code, considerable sequence variation is possible among these polynucleotide
 molecules. SEQ ID NO:3 is a degenerate DNA sequence that encompasses all DNAs
 that encode the znssp6 polypeptide of SEQ ID NO:2. Those skilled in the art will
 recognize that the degenerate sequence of SEQ ID NO:3 also provides all RNA
 sequences encoding SEQ ID NO:2 by substituting U for T. Thus, znssp6 polypeptide-
 20 encoding polynucleotides comprising nucleotide 1 to nucleotide 1134 of SEQ ID NO:3
 and their RNA equivalents are contemplated by the present invention. Table 1 sets
 forth the one-letter codes used within SEQ ID NO:3 to denote degenerate nucleotide
 positions. "Resolutions" are the nucleotides denoted by a code letter. "Complement"
 indicates the code for the complementary nucleotide(s). For example, the code Y
 25 denotes either C or T, and its complement R denotes A or G, A being complementary to
 T, and G being complementary to C.

TABLE 1

Nucleotide	Resolution	Complement	Resolution
A	A	T	T
C	C	G	G
G	G	C	C
T	T	A	A
R	A G	Y	C T
Y	C T	R	A G
M	A C	K	G T
K	G T	M	A C
S	C G	S	C G
W	A T	W	A T
H	A C T	D	A G T
B	C G T	V	A C G
V	A C G	B	C G T
D	A G T	H	A C T
N	A C G T	N	A C G T

The degenerate codons used in SEQ ID NO:3, encompassing all possible
5 codons for a given amino acid, are set forth in Table 2.

TABLE 2

Amino Acid	One Letter Code	Codons	Degenerate Codon
Cys	C	TGC TGT	TGY
Ser	S	AGC AGT TCA TCC TCG TCT	WSN
Thr	T	ACA ACC ACG ACT	ACN
Pro	P	CCA CCC CCG CCT	CCN
Ala	A	GCA GCC GCG GCT	GCN
Gly	G	GGA GGC GGG GGT	GGN
Asn	N	AAC AAT	AAY
Asp	D	GAC GAT	GAY
Glu	E	GAA GAG	GAR
Gln	Q	CAA CAG	CAR
His	H	CAC CAT	CAY
Arg	R	AGA AGG CGA CGC CGG CGT	MGN
Lys	K	AAA AAG	AAR
Met	M	ATG	ATG
Ile	I	ATA ATC ATT	ATH
Leu	L	CTA CTC CTG CTT TTA TTG	YTN
Val	V	GTA GTC GTG GTT	GTN
Phe	F	TTC TTT	TTY
Tyr	Y	TAC TAT	TAY
Trp	W	TGG	TGG
Ter	.	TAA TAG TGA	TRR
Asn Asp	B		RAY
Glu Gln	Z		SAR
Any	X		NNN

One of ordinary skill in the art will appreciate that some ambiguity is introduced in determining a degenerate codon, representative of all possible codons encoding each amino acid. For example, the degenerate codon for serine (WSN) can, in some circumstances, encode arginine (AGR), and the degenerate codon for arginine (MGN) can, in some circumstances, encode serine (AGY). A similar relationship exists between codons encoding phenylalanine and leucine. Thus, some polynucleotides encompassed by the degenerate sequence may encode variant amino acid sequences, but one of ordinary skill in the art can easily identify such variant sequences by reference to the amino acid sequence of SEQ ID NO:2. Variant sequences can be readily tested for functionality as described herein.

One of ordinary skill in the art will also appreciate that different species can exhibit "preferential codon usage." In general, see, Grantham, et al., Nuc. Acids Res. 8:1893-912, 1980; Haas, et al. Curr. Biol. 6:315-24, 1996; Wain-Hobson, et al., Gene 13:355-64, 1981; Grosjean and Fiers, Gene 18:199-209, 1982; Holm, Nuc. Acids Res. 14:3075-87, 1986; Ikemura, J. Mol. Biol. 158:573-97, 1982. As used herein, the term "preferential codon usage" or "preferential codons" is a term of art referring to protein translation codons that are most frequently used in cells of a certain species, thus favoring one or a few representatives of the possible codons encoding each amino acid (See Table 2). For example, the amino acid Threonine (Thr) may be encoded by ACA, ACC, ACG, or ACT, but in mammalian cells ACC is the most commonly used codon; in other species, for example, insect cells, yeast, viruses or bacteria, different Thr codons may be preferential. Preferential codons for a particular species can be introduced into the polynucleotides of the present invention by a variety of methods known in the art. Introduction of preferential codon sequences into recombinant DNA can, for example, enhance production of the protein by making protein translation more efficient within a particular cell type or species. Therefore, the degenerate codon sequence disclosed in SEQ ID NO:3 serves as a template for optimizing expression of polynucleotides in various cell types and species commonly used in the art and disclosed herein. Sequences containing preferential codons can be tested and optimized for expression in various species, and tested for functionality as disclosed herein.

Within preferred embodiments of the invention the isolated polynucleotides will hybridize to similar sized regions of SEQ ID NO:1, other polynucleotide probes, primers, fragments and sequences recited herein or sequences complementary thereto. Polynucleotide hybridization is well known in the art and
 5 widely used for many applications, see for example, Sambrook et al., Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor, NY, 1989; Ausubel et al., eds., Current Protocols in Molecular Biology, John Wiley and Sons, Inc., NY, 1987; Berger and Kimmel, eds., Guide to Molecular Cloning Techniques, Methods in Enzymology, volume 152, 1987 and Wetmur, Crit. Rev. Biochem. Mol.
 10 Biol. 26:227-59, 1990. Polynucleotide hybridization exploits the ability of single stranded complementary sequences to form a double helix hybrid. Such hybrids include DNA-DNA, RNA-RNA and DNA-RNA.

Hybridization will occur between sequences which contain some degree of complementarity. Hybrids can tolerate mismatched base pairs in the double helix,
 15 but the stability of the hybrid is influenced by the degree of mismatch. The T_m of the mismatched hybrid decreases by 1°C for every 1-1.5% base pair mismatch. Varying the stringency of the hybridization conditions allows control over the degree of mismatch that will be present in the hybrid. The degree of stringency increases as the hybridization temperature increases and the ionic strength of the hybridization buffer
 20 decreases. Hybridization buffers generally contain blocking agents such as Denhardt's solution (Sigma Chemical Co., St. Louis, Mo.), denatured salmon sperm DNA, milk powders (BLOTTO), heparin or SDS, and a Na^+ source, such as SSC (1X SSC: 0.15 M NaCl, 15 mM sodium citrate) or SSPE (1X SSPE: 1.8 M NaCl, 10 mM NaH_2PO_4 , 1 mM EDTA, pH 7.7). By decreasing the ionic concentration of the buffer, the stability
 25 of the hybrid is increased. Typically, hybridization buffers contain from between 10 mM-1 M Na^+ . Premixed hybridization solutions are also available from commercial sources such as Clontech Laboratories (Palo Alto, CA) and Promega Corporation (Madison, WI) for use according to manufacturer's instruction. Addition of destabilizing or denaturing agents such as formamide, tetralkylammonium salts,
 30 guanidinium cations or thiocyanate cations to the hybridization solution will alter the T_m of a hybrid. Typically, formamide is used at a concentration of up to 50% to allow

incubations to be carried out at more convenient and lower temperatures. Formamide also acts to reduce non-specific background when using RNA probes.

Stringent hybridization conditions encompass temperatures of about 5-25°C below the thermal melting point T_m of the hybrid and a hybridization buffer having up to 1 M Na^+ . Higher degrees of stringency at lower temperatures can be achieved with the addition of formamide which reduces the T_m of the hybrid about 1°C for each 1% formamide in the buffer solution. Generally, such stringent conditions include temperatures of 20-70°C and a hybridization buffer containing 5X to 6X SSC and 0-50% formamide. A higher degree of stringency can be achieved at temperatures of from 40-70°C with a hybridization buffer having 3X to 4X SSC and from 0-50% formamide. Highly stringent conditions typically encompass temperatures of 42-70°C with a hybridization buffer having up to 2X SSC and 0-50% formamide. Different degrees of stringency can be used during hybridization and washing to achieve maximum specific binding to the target sequence. Typically, the washes following hybridization are performed at increasing degrees of stringency to remove non-hybridized polynucleotide probes from hybridized complexes.

The above conditions are meant to serve as a guide and it is well within the abilities of one skilled in the art to adapt these conditions for use with a particular polypeptide hybrid. The T_m for a specific target sequence is the temperature (under defined conditions) at which 50% of the target sequence will hybridize to a perfectly matched probe sequence. Those conditions that influence the T_m include, the size and base pair content of the polynucleotide probe, the ionic strength of the hybridization solution, and the presence of destabilizing agents in the hybridization solution. Numerous equations for calculating T_m are known in the art, see for example (Sambrook et al., *ibid.*; Ausubel et al., *ibid.*; Berger and Kimmel, *ibid.* and Wetmur, *ibid.*) and are specific for DNA, RNA and DNA-RNA hybrids and polynucleotide probe sequences of varying length. Sequence analysis software such as Oligo 4.0 and Primer Premier, as well as sites on the Internet, are available tools for analyzing a given sequence and calculating T_m based on user defined criteria. Such programs can also analyze a given sequence under defined conditions and suggest suitable probe

sequences. Typically, hybridization of longer polynucleotide sequences, >50 bp, is done at temperatures of about 20-25°C below the calculated T_m . For smaller probes, <50 bp, hybridization is typically carried out at the T_m or 5-10°C below. This allows for the maximum rate of hybridization for DNA-DNA and DNA-RNA hybrids.

5 As previously noted, the isolated polynucleotides of the present invention include DNA and RNA. Methods for preparing DNA and RNA are well known in the art. In general, RNA is isolated from a tissue or cell that produces large amounts of znssp6 RNA. Such tissues and cells are identified by Northern blotting (Thomas, Proc. Natl. Acad. Sci. USA 77:5201, 1980), and include brain, kidney, and
10 testis, spinal cord, colon, prostate, stomach, ovary, pancreas, pituitary gland, adrenal gland, salivary gland, mammary gland, liver, small intestine, spleen, thymus, peripheral leukocyte, lymph node, bone marrow, lung, trachea, placenta, fetal spleen and fetal lung.

Total RNA can be prepared using guanidine isothiocyanate extraction
15 followed by isolation by centrifugation in a CsCl gradient (Chirgwin et al., Biochemistry 18:52-94, 1979). Poly (A)⁺ RNA is prepared from total RNA using the method of Aviv and Leder (Proc. Natl. Acad. Sci. USA 69:1408-12, 1972). Complementary DNA (cDNA) is prepared from poly(A)⁺ RNA using known methods. In the alternative, genomic DNA can be isolated. Polynucleotides encoding znssp6
20 polypeptides are then identified and isolated by, for example, hybridization or PCR.

A full-length clone encoding znssp6 can be obtained by conventional cloning procedures. Complementary DNA (cDNA) clones are preferred, although for some applications (e.g., expression in transgenic animals) it may be preferable to use a genomic clone, or to modify a cDNA clone to include at least one genomic intron.
25 Methods for preparing cDNA and genomic clones are well known and within the level of ordinary skill in the art, and include the use of the sequence disclosed herein, or parts thereof, for probing or priming a library. Expression libraries can be probed with antibodies to znssp6, or fragments thereof, or other specific binding partners.

The invention also provides isolated and purified znssp6 polynucleotide
30 probes. Such polynucleotide probes can be RNA or DNA. DNA can be either cDNA

or genomic DNA. Polynucleotide probes are single or double-stranded DNA or RNA, generally synthetic oligonucleotides, but may be generated from cloned cDNA or genomic sequences and will generally comprise at least 16 nucleotides, more often from 17 nucleotides to 25 or more nucleotides, sometimes 40 to 60 nucleotides, and in some instances a substantial portion, domain or even the entire znssp6 gene or cDNA. The synthetic oligonucleotides of the present invention have at least 75% identity to a representative znssp6 DNA sequence (SEQ ID NO:1 or 3) or their complements. The invention also provides oligonucleotide probes or primers comprising at least 14 contiguous nucleotides of a polynucleotide of SEQ ID NOs: 1 or 3 or a sequence complementary to SEQ ID NOs: 1 or 3.

Regions from which to construct probes include the 5' and/or 3' coding sequences, the anti-complementary molecule-binding regions, the stem domain, and the hydrophobic domain, and the like. Techniques for developing polynucleotide probes and hybridization techniques are known in the art, see for example, Ausubel et al., eds., Current Protocols in Molecular Biology, John Wiley and Sons, Inc., NY, 1991. For use as probes, the molecules can be labeled to provide a detectable signal, such as with an enzyme, biotin, a radionuclide, fluorophore, chemiluminescer, paramagnetic particle and the like, which are commercially available from many sources, such as Molecular Probes, Inc., Eugene, OR, and Amersham Corp., Arlington Heights, IL, using techniques that are well known in the art. Such probes can also be used in hybridizations to detect the presence or quantify the amount of znssp6 gene or mRNA transcript in a sample. Znssp6 polynucleotide probes could be used to hybridize to DNA or RNA targets for diagnostic purposes, using such techniques such as fluorescent *in situ* hybridization (FISH) or immunohistochemistry. Polynucleotide probes can be used to identify genes encoding znssp6-like proteins. For example, znssp6 polynucleotides can be used as primers and/or templates in PCR reactions to identify other novel members of the UDP-glycosyltransferase family. Such probes can also be used to screen libraries for related sequences encoding novel UDP-glycosyltransferases. Such screening would be carried out under conditions of low stringency which would allow identification of sequences which are substantially homologous, but not requiring complete homology to the probe sequence. Such methods and conditions are well

known in the art, see, for example, Sambrook et al., Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor, NY, 1989. Such low stringency conditions could include hybridization temperatures less than 42°C, formamide concentrations of less than 50% and moderate to low concentrations of salt. Libraries
 5 may be made of genomic DNA or cDNA. Polynucleotide probes are also useful for Southern, Northern, or dot blots, colony and plaque hybridization and *in situ* hybridization. Mixtures of different znssp6 polynucleotide probes can be prepared which would increase sensitivity or the detection of low copy number targets, in screening systems.

10 The polynucleotides of the present invention can also be synthesized using DNA synthesizers. Currently the method of choice is the phosphoramidite method. If chemically synthesized double stranded DNA is required for an application such as the synthesis of a gene or a gene fragment, then each complementary strand is made separately. The production of short genes (60 to 80 bp) is technically
 15 straightforward and can be accomplished by synthesizing the complementary strands and then annealing them. For the production of longer genes (>300 bp), however, special strategies must be invoked, because the coupling efficiency of each cycle during chemical DNA synthesis is seldom 100%. To overcome this problem, synthetic genes (double-stranded) are assembled in modular form from single-stranded fragments that
 20 are from 20 to 100 nucleotides in length. See Glick and Pasternak, Molecular Biotechnology, Principles & Applications of Recombinant DNA, (ASM Press, Washington, D.C. 1994); Itakura et al., Annu. Rev. Biochem. 53: 323-56, 1984 and Climie et al., Proc. Natl. Acad. Sci. USA 87:633-7, 1990.

The present invention further provides counterpart polypeptides and
 25 polynucleotides from other species (orthologs). These species include, but are not limited to mammalian, avian, amphibian, reptile, fish, insect and other vertebrate and invertebrate species. Of particular interest are znssp6 polypeptides from other mammalian species, including murine, porcine, ovine, bovine, canine, feline, equine, and other primate polypeptides. Orthologs of human znssp6 can be cloned using
 30 information and compositions provided by the present invention in combination with conventional cloning techniques. For example, a cDNA can be cloned using mRNA

obtained from a tissue or cell type that expresses znssp6 as disclosed herein. Suitable sources of mRNA can be identified by probing Northern blots with probes designed from the sequences disclosed herein. A library is then prepared from mRNA of a positive tissue or cell line. A znssp6-encoding cDNA can then be isolated by a variety of methods, such as by probing with a complete or partial human cDNA or with one or more sets of degenerate probes based on the disclosed sequences. A cDNA can also be cloned using the polymerase chain reaction, or PCR (Mullis, U.S. Patent No. 4,683,202), using primers designed from the representative human znssp6 sequence disclosed herein. Within an additional method, the cDNA library can be used to transform or transfect host cells, and expression of the cDNA of interest can be detected with an antibody to znssp6 polypeptide. Similar techniques can also be applied to the isolation of genomic clones.

Those skilled in the art will recognize that the sequence disclosed in SEQ ID NO:1 represents a single allele of human znssp6 and that allelic variation and alternative splicing are expected to occur. Allelic variants of this sequence can be cloned by probing cDNA or genomic libraries from different individuals according to standard procedures. Allelic variants of the DNA sequence shown in SEQ ID NO:1, including those containing silent mutations and those in which mutations result in amino acid sequence changes, are within the scope of the present invention, as are proteins which are allelic variants of SEQ ID NO:2. cDNAs generated from alternatively spliced mRNAs, which retain the properties of the znssp6 polypeptide are included within the scope of the present invention, as are polypeptides encoded by such cDNAs and mRNAs. Allelic variants and splice variants of these sequences can be cloned by probing cDNA or genomic libraries from different individuals or tissues according to standard procedures known in the art.

The present invention also provides isolated znssp6 polypeptides that are substantially homologous to the polypeptides of SEQ ID NO:2 and their orthologs. The term "substantially homologous" is used herein to denote polypeptides having 50%, preferably 60%, more preferably at least 80%, sequence identity to the sequences shown in SEQ ID NO:2 or their orthologs. Such polypeptides will more preferably be at least 90% identical, and most preferably 95% or more identical to SEQ ID NO:2 or

5

10 Total number of identical matches

[length of the longer sequence plus the
number of gaps introduced into the longer
sequence in order to align the two sequences]

Table 3[illegible]

Sequence identity of polynucleotide molecules is determined by similar methods using a ratio as disclosed above.

Those skilled in the art appreciate that there are many established algorithms available to align two amino acid sequences. The "FASTA" similarity search algorithm of Pearson and Lipman is a suitable protein alignment method for examining the level of identity shared by an amino acid sequence disclosed herein and the amino acid sequence of a putative variant znssp6. The FASTA algorithm is described by Pearson and Lipman, Proc. Nat'l Acad. Sci. USA 85:2444 (1988), and by Pearson, Meth. Enzymol. 183:63 (1990).

Briefly, FASTA first characterizes sequence similarity by identifying regions shared by the query sequence (*e.g.*, SEQ ID NO:2) and a test sequence that have either the highest density of identities (if the ktup variable is 1) or pairs of identities (if ktup=2), without considering conservative amino acid substitutions, insertions, or deletions. The ten regions with the highest density of identities are then rescored by comparing the similarity of all paired amino acids using an amino acid substitution matrix, and the ends of the regions are "trimmed" to include only those residues that contribute to the highest score. If there are several regions with scores greater than the "cutoff" value (calculated by a predetermined formula based upon the length of the sequence and the ktup value), then the trimmed initial regions are examined to determine whether the regions can be joined to form an approximate alignment with gaps. Finally, the highest scoring regions of the two amino acid sequences are aligned using a modification of the Needleman-Wunsch-Sellers algorithm (Needleman and Wunsch, J. Mol. Biol. 48:444 (1970); Sellers, SIAM J. Appl. Math. 26:787 (1974)), which allows for amino acid insertions and deletions. Illustrative parameters for FASTA analysis are: ktup=1, gap opening penalty=10, gap extension penalty=1, and substitution matrix=BLOSUM62. These parameters can be introduced into a FASTA program by modifying the scoring matrix file ("SMATRIX"), as explained in Appendix 2 of Pearson, Meth. Enzymol. 183:63 (1990).

FASTA can also be used to determine the sequence identity of nucleic acid molecules using a ratio as disclosed above. For nucleotide sequence comparisons, the ktup value can range between one to six, preferably from four to six.

The present invention includes nucleic acid molecules that encode a polypeptide having one or more conservative amino acid changes, compared with the amino acid sequence of SEQ ID NO:2. The BLOSUM62 table is an amino acid substitution matrix derived from about 2,000 local multiple alignments of protein sequence segments, representing highly conserved regions of more than 500 groups of related proteins (Henikoff and Henikoff, Proc. Nat'l Acad. Sci. USA 89:10915 (1992)). Accordingly, the BLOSUM62 substitution frequencies can be used to define conservative amino acid substitutions that may be introduced into the amino acid sequences of the present invention. As used herein, the language "conservative amino acid substitution" refers to a substitution represented by a BLOSUM62 value of greater than -1. For example, an amino acid substitution is conservative if the substitution is characterized by a BLOSUM62 value of 0, 1, 2, or 3. Conservative amino acid substitutions are characterized by a BLOSUM62 value of at least 1 (e.g., 1, 2 or 3), while more conservative amino acid substitutions are characterized by a BLOSUM62 value of at least 2 (e.g., 2 or 3).

Variant znssp6 polypeptides or substantially homologous znssp6 polypeptides are characterized as having one or more amino acid substitutions, deletions or additions. These changes are preferably of a minor nature, that is conservative amino acid substitutions (see Table 4) and other substitutions that do not significantly affect the folding or activity of the polypeptide; small deletions, typically of one to about 30 amino acids; and small amino- or carboxyl-terminal extensions, such as an amino-terminal methionine residue, a small linker peptide of up to about 20-25 residues, or an affinity tag. The present invention thus includes polypeptides of from 378 to 410 amino acid residues that comprise a sequence that is at least 70%, preferably at least 80%, and more preferably 90% or more identical to the corresponding region of SEQ ID NO:2. Polypeptides comprising affinity tags can further comprise a proteolytic cleavage site between the znssp6 polypeptide and the affinity tag. Preferred such sites include thrombin cleavage sites and factor Xa cleavage sites.

Table 4
Conservative amino acid substitutions

5	Basic:	arginine
		lysine
		histidine
	Acidic:	glutamic acid
		aspartic acid
	Polar:	glutamine
10		asparagine
	Hydrophobic:	leucine
		isoleucine
		valine
	Aromatic:	phenylalanine
		tryptophan
15		tyrosine
	Small:	glycine
		alanine
		serine
		threonine
		methionine

The present invention further provides a variety of other polypeptide fusions and related multimeric proteins comprising one or more polypeptide fusions.

25 For example, a znssp6 polypeptide can be prepared as a fusion to a dimerizing protein as disclosed in U.S. Patents Nos. 5,155,027 and 5,567,584. Preferred dimerizing proteins in this regard include immunoglobulin heavy chain constant region domains. Immunoglobulin-znssp6 polypeptide fusions can be expressed in genetically engineered cells to produce a variety of multimeric znssp6 analogs wherein the Fc portions are

30 disulfide bonded to each other and two non-Ig polypeptides are arrayed in closed proximity to each other. Fusions of this type can be used to evaluate specific

donor/acceptor molecules, affinity purify ligands, or use as an *in vitro* assay tool. This fusion can also be used to determine the homodimerization potential for znssp6. For use in assays, the chimeras are bound to a support via the F_C region and used in an ELISA format.

5 Auxiliary domains can be fused to znssp6 polypeptides to target them to specific cells, tissues, or macromolecules (e.g., brain, kidney, and testis, for example). For example, a protease, or ablation antibody polypeptide or protein could be targeted to a predetermined cell type by fusing a said protease, or ablation antibody polypeptide to a ligand that specifically binds to a receptor or receptor-like complementary
10 molecule on the surface of the target cell, such as, brain, kidney, and testis. In this way, polypeptides and proteins can be targeted for therapeutic or diagnostic purposes. Such beta-1,3-galactosyltransferase polypeptides can be fused to two or more moieties, such as an affinity tag for purification and a targeting domain. Polypeptide fusions can also comprise one or more cleavage sites, particularly between domains. See, Tuan et al.,
15 Connective Tissue Research 34:1-9, 1996.

Polypeptide fusions of the present invention will generally contain not more than about 1,700 amino acid residues, not more than about 1,200 residues, or not more than about 1,000 residues, and will in many cases be considerably smaller. For example, residues of znssp6 polypeptide can be fused to *E. coli* β -galactosidase (1,021
20 residues; see Casadaban et al., J. Bacteriol. 143:971-980, 1980), a 10-residue spacer, and a 4-residue factor Xa cleavage site. In a second example, residues of znssp6 polypeptide can be fused to maltose binding protein (approximately 370 residues), a 4-residue cleavage site, and a 6-residue polyhistidine tag.

Some proteins in the β 3Gal-T family been shown to be expressed
25 intracellularly and are involved in intracellular glycoprotein and glycolipid processing. Other members of this family have been shown to be extracellularly expressed and are involved in glycoprotein and glycolipid processing (such as in the case of the Tn antigen). Other members of the family are expressed extracellularly and are involved in cell-cell interactions and intracellular signaling. Thus, molecules of the present
30 invention can function as an enzyme both intracellularly and extracellularly, in which case its anti-complementary molecule is a substrate. Additionally, molecules of the

present invention can function extracellularly and modulate cell-cell interactions. The extracellular binding of znssp6 to its anti-complementary molecule can cause a cellular event in the cell that is expressing it (i.e. znssp6 acts as a receptor or receptor-like molecule), or in the cell expressing the anti-complementary molecule to which it binds (i.e., znssp6 acts as a ligand). Additionally, znssp6 can function extracellularly as a soluble enzyme, ligand, receptor or receptor-like molecule. Similarly, as an extracellularly expressed znssp6 enzyme, the processing of its anti-complementary substrate can result in a cellular response (similar to intracellular signaling) in the cell expressing the substrate. Also as an extracellularly expressed protein, znssp6 can function to form a "bridge" between cells maintaining their proximity to each other. Thus, for the purposes of this application, znssp6 is referred to as a complementary molecule and its cognate binding partner is referred to as an anti-complementary molecule.

The invention also provides soluble znssp6 polypeptides, used to form fusion or chimeric proteins with human Ig, as His-tagged proteins, or FLAGTM-tagged proteins. One such construct is comprises residues 50 to 370 of SEQ ID NO:2, fused to human Ig. znssp6 or znssp6-Ig chimeric proteins are used, for example, to identify the znssp6 anti-complementary molecule, including the natural anti-complementary molecule, as well as agonists and antagonists of the natural anti-complementary molecule. Using labeled soluble znssp6, cells expressing the anti-complementary molecule are identified by fluorescence immunocytometry or immunohistochemistry. The soluble fusion proteins or soluble Ig fusion protein is useful in studying the distribution of the anti-complementary molecule on tissues or specific cell lineages, and to provide insight into complementary molecule-anti-complementary molecule biology.

In an alternative approach, a soluble znssp6 extracellular anti-complementary molecule-binding region can be expressed as a chimera with immunoglobulin heavy chain constant regions, typically an F_C fragment, which contains two constant region domains and a hinge region, but lacks the variable region. Such fusions are typically secreted as multimeric molecules, wherein the Fc portions are disulfide bonded to each other and two znssp6 polypeptides are arrayed in close proximity to each other. Fusions of this type can be used to affinity purify the cognate

anti-complementary molecule from solution, as an *in vitro* assay tool, to block signals *in vitro* by specifically titrating out anti-complementary molecule, and as antagonists *in vivo* by administering them to block the anti-complementary molecule stimulation. To purify anti-complementary molecule, a znssp6-Ig fusion protein (chimera) is added to a sample containing the anti-complementary molecule under conditions that facilitate complementary molecule-anti-complementary molecule binding (typically near-physiological temperature, pH, and ionic strength). The chimera- anti-complementary molecule complex is then separated by the mixture using protein A, which is immobilized on a solid support (e.g., insoluble resin beads). The anti-complementary molecule is then eluted using conventional chemical techniques, such as with a salt or pH gradient. In the alternative, the chimera itself can be bound to a solid support, with binding and elution carried out as above. For use in assays, the chimeras are bound to a support via the F_C region and used in an ELISA format.

The present invention also includes "functional fragments" of znssp6 polypeptides and nucleic acid molecules encoding such functional fragments. Routine deletion analyses of nucleic acid molecules can be performed to obtain functional fragments of a nucleic acid molecule that encodes a znssp6 polypeptide. As an illustration, DNA molecules having the nucleotide sequence of SEQ ID NO:1 can be digested with *Bal31* nuclease to obtain a series of nested deletions. The fragments are then inserted into expression vectors in proper reading frame, and the expressed polypeptides are isolated and tested for cell-cell interactions, or for the ability to bind anti-znssp6 antibodies. One alternative to exonuclease digestion is to use oligonucleotide-directed mutagenesis to introduce deletions or stop codons to specify production of a desired fragment. Alternatively, particular fragments of an znssp6 gene can be synthesized using the polymerase chain reaction.

Standard methods for identifying functional domains are well-known to those of skill in the art. For example, studies on the truncation at either or both termini of interferons have been summarized by Horisberger and Di Marco, Pharmac. Ther. 66:507 (1995). Moreover, standard techniques for functional analysis of proteins are described by, for example, Treuter *et al.*, Molec. Gen. Genet. 240:113 (1993), Content *et al.*, "Expression and preliminary deletion analysis of the 42 kDa 2-5A synthetase

induced by human interferon," in *Biological Interferon Systems, Proceedings of ISIR-TNO Meeting on Interferon Systems*, Cantell (ed.), pages 65-72 (Nijhoff 1987), Herschman, "The EGF Enzyme," in *Control of Animal Cell Proliferation, Vol. 1*, Boynton *et al.*, (eds.) pages 169-199 (Academic Press 1985), Coumailleau *et al.*, J. Biol. Chem. 270:29270 (1995); Fukunaga *et al.*, J. Biol. Chem. 270:25291 (1995); Yamaguchi *et al.*, Biochem. Pharmacol. 50:1295 (1995), and Meisel *et al.*, Plant Molec. Biol. 30:1 (1996).

The present invention also contemplates functional fragments of an znssp6 gene that has amino acid changes, compared with the amino acid sequence of SEQ ID NO:2. A variant znssp6 gene can be identified on the basis of structure by determining the level of identity with nucleotide and amino acid sequences of SEQ ID NOs:1 and 2, as discussed above. An alternative approach to identifying a variant gene on the basis of structure is to determine whether a nucleic acid molecule encoding a potential variant znssp6 gene can hybridize to a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1, as discussed above.

The proteins of the present invention can also comprise non-naturally occurring amino acid residues. Non-naturally occurring amino acids include, without limitation, *trans*-3-methylproline, 2,4-methanoproline, *cis*-4-hydroxyproline, *trans*-4-hydroxyproline, *N*-methylglycine, *allo*-threonine, methylthreonine, hydroxyethylcysteine, hydroxyethylhomocysteine, nitroglutamine, homoglutamine, pipecolic acid, thiazolidine carboxylic acid, dehydroproline, 3- and 4-methylproline, 3,3-dimethylproline, *tert*-leucine, norvaline, 2-azaphenylalanine, 3-azaphenylalanine, 4-azaphenylalanine, and 4-fluorophenylalanine. Several methods are known in the art for incorporating non-naturally occurring amino acid residues into proteins. For example, an *in vitro* system can be employed wherein nonsense mutations are suppressed using chemically aminoacylated suppressor tRNAs. Methods for synthesizing amino acids and aminoacylating tRNA are known in the art. Transcription and translation of plasmids containing nonsense mutations is carried out in a cell-free system comprising an *E. coli* S30 extract and commercially available enzymes and other reagents. Proteins are purified by chromatography. See, for example, Robertson *et al.*, J. Am. Chem. Soc. 113:2722, 1991; Ellman *et al.*, Methods Enzymol. 202:301, 1991; Chung *et al.*, Science

259:806-9, 1993; and Chung et al., Proc. Natl. Acad. Sci. USA 90:10145-9, 1993). In a second method, translation is carried out in *Xenopus* oocytes by microinjection of mutated mRNA and chemically aminoacylated suppressor tRNAs (Turcatti et al., J. Biol. Chem. 271:19991-8, 1996). Within a third method, *E. coli* cells are cultured in the absence of a natural amino acid that is to be replaced (e.g., phenylalanine) and in the presence of the desired non-naturally occurring amino acid(s) (e.g., 2-azaphenylalanine, 3-azaphenylalanine, 4-azaphenylalanine, or 4-fluorophenylalanine). The non-naturally occurring amino acid is incorporated into the protein in place of its natural counterpart. See, Koide et al., Biochem. 33:7470-6, 1994. Naturally occurring amino acid residues can be converted to non-naturally occurring species by *in vitro* chemical modification. Chemical modification can be combined with site-directed mutagenesis to further expand the range of substitutions (Wynn and Richards, Protein Sci. 2:395-403, 1993).

A limited number of non-conservative amino acids, amino acids that are not encoded by the genetic code, non-naturally occurring amino acids, and unnatural amino acids may be substituted for znssp6 amino acid residues.

Essential amino acids in the polypeptides of the present invention can be identified according to procedures known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis (Cunningham and Wells, Science 244: 1081-5, 1989; Bass et al., Proc. Natl. Acad. Sci. USA 88:4498-502, 1991). In the latter technique, single alanine mutations are introduced at every residue in the molecule, and the resultant mutant molecules are tested for biological activity as disclosed below to identify amino acid residues that are critical to the activity of the molecule. See also, Hilton et al., J. Biol. Chem. 271:4699-708, 1996. Sites of galactosyltransferase, or znssp6 anti-complementary molecule binding activity can also be determined by physical analysis of structure, as determined by such techniques as nuclear magnetic resonance, crystallography, electron diffraction or photoaffinity labeling, in conjunction with mutation of putative contact site amino acids. See, for example, de Vos et al., Science 255:306-12, 1992; Smith et al., J. Mol. Biol. 224:899-904, 1992; Wlodaver et al., FEBS Lett. 309:59-64, 1992. The identities of essential amino acids can also be inferred from analysis of homologies with related galactosyltransferase molecules.

Multiple amino acid substitutions can be made and tested using known methods of mutagenesis and screening, such as those disclosed by Reidhaar-Olson and Sauer (Science 241:53-7, 1988) or Bowie and Sauer (Proc. Natl. Acad. Sci. USA 86:2152-6, 1989). Briefly, these authors disclose methods for simultaneously
 5 randomizing two or more positions in a polypeptide, selecting for functional polypeptide, and then sequencing the mutagenized polypeptides to determine the spectrum of allowable substitutions at each position. Other methods that can be used include phage display (e.g., Lowman et al., Biochem 30:10832-7, 1991; Ladner et al., U.S. Patent No. 5,223,409; Huse, WIPO Publication WO 92/06204) and region-
 10 directed mutagenesis (Derbyshire et al., Gene 46:145, 1986; Ner et al., DNA 7:127, 1988).

Variants of the disclosed znssp6 DNA and polypeptide sequences can be generated through DNA shuffling as disclosed by Stemmer, Nature 370:389-91, 1994, Stemmer, Proc. Natl. Acad. Sci. USA 91:10747-51, 1994 and WIPO Publication WO
 15 97/20078. Briefly, variant DNAs are generated by *in vitro* homologous recombination by random fragmentation of a parent DNA followed by reassembly using PCR, resulting in randomly introduced point mutations. This technique can be modified by using a family of parent DNAs, such as allelic variants or DNAs from different species, to introduce additional variability into the process. Selection or screening for the
 20 desired activity, followed by additional iterations of mutagenesis and assay provides for rapid "evolution" of sequences by selecting for desirable mutations while simultaneously selecting against detrimental changes.

Mutagenesis methods as disclosed herein can be combined with high-throughput, automated screening methods to detect activity of cloned, mutagenized
 25 polypeptides in host cells. Mutagenized DNA molecules that encode active polypeptides (e.g., galactosyltransferase activity as evidenced by glycoprotein synthesis, or cell-cell interactions) can be recovered from the host cells and rapidly sequenced using modern equipment. These methods allow the rapid determination of the importance of individual amino acid residues in a polypeptide of interest, and can
 30 be applied to polypeptides of unknown structure.

Regardless of the particular nucleotide sequence of a variant znssp6 gene, the gene encodes a polypeptide that is characterized by its anti-complementary molecule binding activity, or by the ability to bind specifically to an anti-znssp6 antibody. More specifically, variant znssp6 genes encode polypeptides which exhibit
 5 greater than 75, 80, or 90%, of the activity of polypeptide encoded by the human znssp6 gene described herein.

Using the methods discussed herein, one of ordinary skill in the art can identify and/or prepare a variety of polypeptide fragments or variants of SEQ ID NO:2 or that retain the galactosyltransferase properties, or cell-cell interactions of the wild-
 10 type znssp6 protein. Such polypeptides may include additional amino acids from, for example, an extracellular anti-complementary molecule-binding domain of another member of the galactosyltransferase family as well as part or all of the transmembrane and intracellular domains. Additionally fragments of znssp6 may include additional amino acids from the catalytic site of the galactosyltransferase domains of other family
 15 members. Additional amino acids from affinity tags and the like may also be included.

For any znssp6 polypeptide, including variants and fusion proteins, one of ordinary skill in the art can readily generate a fully degenerate polynucleotide sequence encoding that variant using the information set forth in Tables 1 and 2 above.

The znssp6 polypeptides of the present invention, including full-length
 20 polypeptides, biologically active fragments, and fusion polypeptides, can be produced in genetically engineered host cells according to conventional techniques. Suitable host cells are those cell types that can be transformed or transfected with exogenous DNA and grown in culture, and include bacteria, fungal cells, and cultured higher eukaryotic cells. Eukaryotic cells, particularly cultured cells of multicellular organisms, are
 25 preferred. Techniques for manipulating cloned DNA molecules and introducing exogenous DNA into a variety of host cells are disclosed by Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989, and Ausubel et al., eds., *Current Protocols in Molecular Biology*, John Wiley and Sons, Inc., NY, 1987.

30 In general, a DNA sequence encoding a znssp6 polypeptide is operably linked to other genetic elements required for its expression, generally including a

transcription promoter and terminator, within an expression vector. The vector will also commonly contain one or more selectable markers and one or more origins of replication, although those skilled in the art will recognize that within certain systems selectable markers may be provided on separate vectors, and replication of the exogenous DNA may be provided by integration into the host cell genome. Selection of promoters, terminators, selectable markers, vectors and other elements is a matter of routine design within the level of ordinary skill in the art. Many such elements are described in the literature and are available through commercial suppliers.

To direct a znssp6 polypeptide into the secretory pathway of a host cell, a secretory signal sequence (also known as a leader sequence, prepro sequence or pre sequence) is provided in the expression vector. The secretory signal sequence may be that of znssp6, or may be derived from another secreted protein (e.g., t-PA) or synthesized *de novo*. The secretory signal sequence is operably linked to the znssp6 DNA sequence, i.e., the two sequences are joined in the correct reading frame and positioned to direct the newly synthesized polypeptide into the secretory pathway of the host cell. Secretory signal sequences are commonly positioned 5' to the DNA sequence encoding the polypeptide of interest, although certain secretory signal sequences may be positioned elsewhere in the DNA sequence of interest (see, e.g., Welch et al., U.S. Patent No. 5,037,743; Holland et al., U.S. Patent No. 5,143,830).

The native secretory signal sequence of the polypeptides of the present invention is used to direct other polypeptides into the secretory pathway. The present invention provides for such fusion polypeptides. A signal fusion polypeptide can be made wherein a secretory signal sequence derived from a znssp6 polypeptide is operably linked to another polypeptide using methods known in the art and disclosed herein. The secretory signal sequence contained in the fusion polypeptides of the present invention is preferably fused amino-terminally to an additional peptide to direct the additional peptide into the secretory pathway. Such constructs have numerous applications known in the art. For example, these novel secretory signal sequence fusion constructs can direct the secretion of an active component of a normally non-secreted protein, such as a receptor. Such fusions may be used *in vivo* or *in vitro* to direct peptides through the secretory pathway.

Cultured mammalian cells are suitable hosts within the present invention. Methods for introducing exogenous DNA into mammalian host cells include calcium phosphate-mediated transfection (Wigler et al., Cell 14:725, 1978; Corsaro and Pearson, Somatic Cell Genetics 7:603, 1981; Graham and Van der Eb, Virology 52:456, 1973), electroporation (Neumann et al., EMBO J. 1:841-5, 1982), DEAE-dextran mediated transfection (Ausubel et al., *ibid.*), and liposome-mediated transfection (Hawley-Nelson et al., Focus 15:73, 1993; Ciccarone et al., Focus 15:80, 1993, and viral vectors (Miller and Rosman, BioTechniques 7:980-90, 1989; Wang and Finer, Nature Med. 2:714-6, 1996). The production of recombinant polypeptides in cultured mammalian cells is disclosed, for example, by Levinson et al., U.S. Patent No. 4,713,339; Hagen et al., U.S. Patent No. 4,784,950; Palmiter et al., U.S. Patent No. 4,579,821; and Ringold, U.S. Patent No. 4,656,134. Suitable cultured mammalian cells include the COS-1 (ATCC No. CRL 1650), COS-7 (ATCC No. CRL 1651), BHK (ATCC No. CRL 1632), BHK 570 (ATCC No. CRL 10314), 293 (ATCC No. CRL 1573; Graham et al., J. Gen. Virol. 36:59-72, 1977) and Chinese hamster ovary (e.g. CHO-K1; ATCC No. CCL 61) cell lines. Additional suitable cell lines are known in the art and available from public depositories such as the American Type Culture Collection, Rockville, Maryland. In general, strong transcription promoters are preferred, such as promoters from SV-40 or cytomegalovirus. See, e.g., U.S. Patent No. 4,956,288. Other suitable promoters include those from metallothionein genes (U.S. Patent Nos. 4,579,821 and 4,601,978) and the adenovirus major late promoter.

Drug selection is generally used to select for cultured mammalian cells into which foreign DNA has been inserted. Such cells are commonly referred to as "transfectants". Cells that have been cultured in the presence of the selective agent and are able to pass the gene of interest to their progeny are referred to as "stable transfectants." A preferred selectable marker is a gene encoding resistance to the antibiotic neomycin. Selection is carried out in the presence of a neomycin-type drug, such as G-418 or the like. Selection systems can also be used to increase the expression level of the gene of interest, a process referred to as "amplification." Amplification is carried out by culturing transfectants in the presence of a low level of the selective agent and then increasing the amount of selective agent to select for cells that produce

high levels of the products of the introduced genes. A preferred amplifiable selectable marker is dihydrofolate reductase, which confers resistance to methotrexate. Other drug resistance genes (e.g. hygromycin resistance, multi-drug resistance, puromycin acetyltransferase) can also be used. Alternative markers that introduce an altered phenotype, such as green fluorescent protein, or cell surface proteins such as CD4, CD8, Class I MHC, placental alkaline phosphatase may be used to sort transfected cells from untransfected cells by such means as FACS sorting or magnetic bead separation technology.

Other higher eukaryotic cells can also be used as hosts, including plant cells, insect cells and avian cells. The use of *Agrobacterium rhizogenes* as a vector for expressing genes in plant cells has been reviewed by Sinkar et al., J. Biosci. (Bangalore) 11:47-58, 1987. Transformation of insect cells and production of foreign polypeptides therein is disclosed by Guarino et al., U.S. Patent No. 5,162,222 and WIPO publication WO 94/06463. Insect cells can be infected with recombinant baculovirus, commonly derived from *Autographa californica nuclear polyhedrosis virus* (AcNPV). See, King, L.A. and Possee, R.D., The Baculovirus Expression System: A Laboratory Guide, London, Chapman & Hall; O'Reilly, D.R. et al., Baculovirus Expression Vectors: A Laboratory Manual, New York, Oxford University Press., 1994; and, Richardson, C. D., Ed., Baculovirus Expression Protocols. Methods in Molecular Biology, Totowa, NJ, Humana Press, 1995. A second method of making recombinant znssp6 baculovirus utilizes a transposon-based system described by Luckow (Luckow, V.A, et al., J. Virol. 67:4566-79, 1993). This system, which utilizes transfer vectors, is sold in the Bac-to-Bac™ kit (Life Technologies, Rockville, MD). This system utilizes a transfer vector, pFastBac1™ (Life Technologies) containing a Tn7 transposon to move the DNA encoding the znssp6 polypeptide into a baculovirus genome maintained in *E. coli* as a large plasmid called a "bacmid." The pFastBac1™ transfer vector utilizes the AcNPV polyhedrin promoter to drive the expression of the gene of interest, in this case znssp6. However, pFastBac1™ can be modified to a considerable degree. The polyhedrin promoter can be removed and substituted with the baculovirus basic protein promoter (also known as P_{cor}, p6.9 or MP promoter) which is expressed earlier in the baculovirus infection, and has been shown to be advantageous

for expressing secreted proteins. See, Hill-Perkins, M.S. and Possee, R.D., J. Gen. Virol. 71:971-6, 1990; Bonning, B.C. et al., J. Gen. Virol. 75:1551-6, 1994; and, Chazenbalk, G.D., and Rapoport, B., J. Biol. Chem. 270:1543-9, 1995. In such transfer vector constructs, a short or long version of the basic protein promoter can be used.

5 Moreover, transfer vectors can be constructed which replace the native znssp6 secretory signal sequences with secretory signal sequences derived from insect proteins. For example, a secretory signal sequence from Ecdysteroid Glucosyltransferase (EGT), honey bee Melittin (Invitrogen, Carlsbad, CA), or baculovirus gp67 (PharMingen, San Diego, CA) can be used in constructs to replace the native znssp6 secretory signal
10 sequence. In addition, transfer vectors can include an in-frame fusion with DNA encoding an epitope tag at the C- or N-terminus of the expressed znssp6 polypeptide, for example, a Glu-Glu epitope tag (Grussenmeyer, T. et al., Proc. Natl. Acad. Sci. 82:7952-4, 1985). Using a technique known in the art, a transfer vector containing znssp6 is transformed into E. coli, and screened for bacmids which contain an
15 interrupted lacZ gene indicative of recombinant baculovirus. The bacmid DNA containing the recombinant baculovirus genome is isolated, using common techniques, and used to transfect *Spodoptera frugiperda* cells, e.g. Sf9 cells. Recombinant virus that expresses znssp6 is subsequently produced. Recombinant viral stocks are made by methods commonly used in the art.

20 The recombinant virus is used to infect host cells, typically a cell line derived from the fall armyworm, *Spodoptera frugiperda*. See, in general, Glick and Pasternak, Molecular Biotechnology: Principles and Applications of Recombinant DNA, ASM Press, Washington, D.C., 1994. Another suitable cell line is the High FiveO™ cell line (Invitrogen) derived from *Trichoplusia ni* (U.S. Patent #5,300,435).
25 Commercially available serum-free media are used to grow and maintain the cells. Suitable media are Sf900 II™ (Life Technologies) or ESF 921™ (Expression Systems) for the Sf9 cells; and Ex-cellO405™ (JRH Biosciences, Lenexa, KS) or Express FiveO™ (Life Technologies) for the *T. ni* cells. The cells are grown up from an inoculation density of approximately $2-5 \times 10^5$ cells to a density of $1-2 \times 10^6$ cells at
30 which time a recombinant viral stock is added at a multiplicity of infection (MOI) of 0.1 to 10, more typically near 3. Procedures used are generally described in available

laboratory manuals (King, L. A. and Possee, R.D., *ibid.*; O'Reilly, D.R. et al., *ibid.*; Richardson, C. D., *ibid.*). Subsequent purification of the znssp6 polypeptide from the supernatant can be achieved using methods described herein.

Fungal cells, including yeast cells, can also be used within the present invention. Yeast species of particular interest in this regard include *Saccharomyces cerevisiae*, *Pichia pastoris*, and *Pichia methanolica*. Methods for transforming *S. cerevisiae* cells with exogenous DNA and producing recombinant polypeptides therefrom are disclosed by, for example, Kawasaki, U.S. Patent No. 4,599,311; Kawasaki et al., U.S. Patent No. 4,931,373; Brake, U.S. Patent No. 4,870,008; Welch et al., U.S. Patent No. 5,037,743; and Murray et al., U.S. Patent No. 4,845,075. Transformed cells are selected by phenotype determined by the selectable marker, commonly drug resistance or the ability to grow in the absence of a particular nutrient (e.g., leucine). A preferred vector system for use in *Saccharomyces cerevisiae* is the *POT1* vector system disclosed by Kawasaki et al. (U.S. Patent No. 4,931,373), which allows transformed cells to be selected by growth in glucose-containing media. Suitable promoters and terminators for use in yeast include those from glycolytic enzyme genes (see, e.g., Kawasaki, U.S. Patent No. 4,599,311; Kingsman et al., U.S. Patent No. 4,615,974; and Bitter, U.S. Patent No. 4,977,092) and alcohol dehydrogenase genes. See also U.S. Patents Nos. 4,990,446; 5,063,154; 5,139,936 and 4,661,454. Transformation systems for other yeasts, including *Hansenula polymorpha*, *Schizosaccharomyces pombe*, *Kluyveromyces lactis*, *Kluyveromyces fragilis*, *Ustilago maydis*, *Pichia pastoris*, *Pichia methanolica*, *Pichia guilliermondii* and *Candida maltosa* are known in the art. See, for example, Gleeson et al., J. Gen. Microbiol. 132:3459-65, 1986 and Cregg, U.S. Patent No. 4,882,279. *Aspergillus* cells may be utilized according to the methods of McKnight et al., U.S. Patent No. 4,935,349. Methods for transforming *Acremonium chrysogenum* are disclosed by Sumino et al., U.S. Patent No. 5,162,228. Methods for transforming *Neurospora* are disclosed by Lambowitz, U.S. Patent No. 4,486,533.

The use of *Pichia methanolica* as host for the production of recombinant proteins is disclosed in WIPO Publications WO 97/17450, WO 97/17451, WO 98/02536, and WO 98/02565. DNA molecules for use in transforming *P. methanolica*

will commonly be prepared as double-stranded, circular plasmids, which are preferably linearized prior to transformation. For polypeptide production in *P. methanolica*, it is preferred that the promoter and terminator in the plasmid be that of a *P. methanolica* gene, such as a *P. methanolica* alcohol utilization gene (*AUG1* or *AUG2*). Other useful promoters include those of the dihydroxyacetone synthase (DHAS), formate dehydrogenase (FMD), and catalase (CAT) genes. To facilitate integration of the DNA into the host chromosome, it is preferred to have the entire expression segment of the plasmid flanked at both ends by host DNA sequences. A preferred selectable marker for use in *Pichia methanolica* is a *P. methanolica ADE2* gene, which encodes phosphoribosyl-5-aminoimidazole carboxylase (AIRC; EC 4.1.1.21), which allows *ade2* host cells to grow in the absence of adenine. For large-scale, industrial processes where it is desirable to minimize the use of methanol, it is preferred to use host cells in which both methanol utilization genes (*AUG1* and *AUG2*) are deleted. For production of secreted proteins, host cells deficient in vacuolar protease genes (*PEP4* and *PRB1*) are preferred. Electroporation is used to facilitate the introduction of a plasmid containing DNA encoding a polypeptide of interest into *P. methanolica* cells. It is preferred to transform *P. methanolica* cells by electroporation using an exponentially decaying, pulsed electric field having a field strength of from 2.5 to 4.5 kV/cm, preferably about 3.75 kV/cm, and a time constant (t) of from 1 to 40 milliseconds, most preferably about 20 milliseconds.

Prokaryotic host cells, including strains of the bacteria *Escherichia coli*, *Bacillus* and other genera are also useful host cells within the present invention. Techniques for transforming these hosts and expressing foreign DNA sequences cloned therein are well known in the art (see, e.g., Sambrook et al., *ibid.*). When expressing a *znssp6* polypeptide in bacteria such as *E. coli*, the polypeptide may be retained in the cytoplasm, typically as insoluble granules, or may be directed to the periplasmic space by a bacterial secretion sequence. In the former case, the cells are lysed, and the granules are recovered and denatured using, for example, guanidine isothiocyanate or urea. The denatured polypeptide can then be refolded and dimerized by diluting the denaturant, such as by dialysis against a solution of urea and a combination of reduced and oxidized glutathione, followed by dialysis against a buffered saline solution. In the

latter case, the polypeptide can be recovered from the periplasmic space in a soluble and functional form by disrupting the cells (by, for example, sonication or osmotic shock) to release the contents of the periplasmic space and recovering the protein, thereby obviating the need for denaturation and refolding.

5 Transformed or transfected host cells are cultured according to conventional procedures in a culture medium containing nutrients and other components required for the growth of the chosen host cells. A variety of suitable media, including defined media and complex media, are known in the art and generally include a carbon source, a nitrogen source, essential amino acids, vitamins and
10 minerals. Media may also contain such components as growth factors or serum, as required. The growth medium will generally select for cells containing the exogenously added DNA by, for example, drug selection or deficiency in an essential nutrient which is complemented by the selectable marker carried on the expression vector or co-transfected into the host cell. *P. methanolica* cells are cultured in a
15 medium comprising adequate sources of carbon, nitrogen and trace nutrients at a temperature of about 25°C to 35°C. Liquid cultures are provided with sufficient aeration by conventional means, such as shaking of small flasks or sparging of fermentors. A preferred culture medium for *P. methanolica* is YEPD (2% D-glucose, 2% Bacto™ Peptone (Difco Laboratories, Detroit, MI), 1% Bacto™ yeast extract (Difco
20 Laboratories), 0.004% adenine and 0.006% L-leucine).

It is preferred to purify the polypeptides of the present invention to ≥80% purity, more preferably to ≥90% purity, even more preferably ≥95% purity, and particularly preferred is a pharmaceutically pure state, that is greater than 99.9% pure with respect to contaminating macromolecules, particularly other proteins and nucleic
25 acids, and free of infectious and pyrogenic agents. Preferably, a purified polypeptide is substantially free of other polypeptides, particularly other polypeptides of animal origin.

Expressed recombinant znssp6 polypeptides (or chimeric znssp6 polypeptides) can be purified using fractionation and/or conventional purification
30 methods and media. Ammonium sulfate precipitation and acid or chaotrope extraction may be used for fractionation of samples. Exemplary purification steps may include

hydroxyapatite, size exclusion, FPLC and reverse-phase high performance liquid chromatography. Suitable chromatographic media include derivatized dextrans, agarose, cellulose, polyacrylamide, specialty silicas, and the like. PEI, DEAE, QAE and Q derivatives are preferred. Exemplary chromatographic media include those media

5 derivatized with phenyl, butyl, or octyl groups, such as Phenyl-Sepharose FF (Pharmacia), Toyopearl butyl 650 (Toso Haas, Montgomeryville, PA), Octyl-Sepharose (Pharmacia) and the like; or polyacrylic resins, such as Amberchrom CG 71 (Toso Haas) and the like. Suitable solid supports include glass beads, silica-based resins, cellulosic resins, agarose beads, cross-linked agarose beads, polystyrene beads, cross-

10 linked polyacrylamide resins and the like that are insoluble under the conditions in which they are to be used. These supports may be modified with reactive groups that allow attachment of proteins by amino groups, carboxyl groups, sulfhydryl groups, hydroxyl groups and/or carbohydrate moieties. Examples of coupling chemistries include cyanogen bromide activation, N-hydroxysuccinimide activation, epoxide

15 activation, sulfhydryl activation, hydrazide activation, and carboxyl and amino derivatives for carbodiimide coupling chemistries. These and other solid media are well known and widely used in the art, and are available from commercial suppliers. Methods for binding receptor and receptor-like complementary polypeptides to support media are well known in the art. Selection of a particular method is a matter of routine

20 design and is determined in part by the properties of the chosen support. See, for example, Affinity Chromatography: Principles & Methods, Pharmacia LKB Biotechnology, Uppsala, Sweden, 1988.

The polypeptides of the present invention can be isolated by a combination of procedures including, but not limited to, anion and cation exchange

25 chromatography, size exclusion, and affinity chromatography. For example, immobilized metal ion adsorption (IMAC) chromatography can be used to purify histidine-rich proteins, including those comprising polyhistidine tags. Briefly, a gel is first charged with divalent metal ions to form a chelate (Sulkowski, Trends in Biochem. 3:1-7, 1985). Histidine-rich proteins will be adsorbed to this matrix with differing

30 affinities, depending upon the metal ion used, and will be eluted by competitive elution, lowering the pH, or use of strong chelating agents. Other methods of purification

include purification of glycosylated proteins by lectin affinity chromatography and ion exchange chromatography (Methods in Enzymol., Vol. 182, "Guide to Protein Purification", M. Deutscher, (ed.), Acad. Press, San Diego, 1990, pp.529-39). Within additional embodiments of the invention, a fusion of the polypeptide of interest and an affinity tag (e.g., maltose-binding protein, an immunoglobulin domain) may be constructed to facilitate purification.

To direct the export of a znssp6 polypeptide from the host cell, the znssp6 DNA is linked to a second DNA segment encoding a secretory peptide, such as a t-PA secretory peptide or a znssp6 secretory peptide. To facilitate purification of the secreted znssp6 polypeptide, a C-terminal extension, such as a poly-histidine tag, substance P, Flag peptide (Hopp et al., Bio/Technology 6:1204-1210, 1988; available from Eastman Kodak Co., New Haven, CT) or another polypeptide or protein for which an antibody or other specific binding agent is available, can be fused to the znssp6 polypeptide.

Moreover, using methods described in the art, polypeptide fusions, or hybrid znssp6 proteins, are constructed using regions or domains of the inventive znssp6 in combination with those of other human galactosyltransferase family proteins (e.g. HSGALT3, HSGALT4, β 3Gal-T2, and β 3Gal-T3, or human homologs to the human ortholog of *Brainiac*), or heterologous proteins (Sambrook et al., *ibid.*, Altschul et al., *ibid.*, Picard, Cur. Opin. Biology, 5:511-5, 1994, and references therein). These methods allow the determination of the biological importance of larger domains or regions in a polypeptide of interest. Such hybrids may alter reaction kinetics, binding, constrict or expand the anti-complementary molecule specificity, or alter tissue and cellular localization of a polypeptide, and can be applied to polypeptides of unknown structure.

Fusion proteins can be prepared by methods known to those skilled in the art by preparing each component of the fusion protein and chemically conjugating them. Alternatively, a polynucleotide encoding both components of the fusion protein in the proper reading frame can be generated using known techniques and expressed by the methods described herein. For example, part or all of a domain(s) conferring a biological function may be swapped between znssp6 of the present invention with the

functionally equivalent domain(s) from another family member, such as the human species ortholog of *Brainiac*, or other galactosyltransferases, etc. Such domains include, but are not limited to, the hydrophobic region thought to be a putative secretory signal sequence or transmembrane domain (residues 26 to 49 of SEQ ID NO:2), the stem or linker domain (residues 50 to 113 of SEQ ID NO:2), and other conserved motifs such as the beta-1,3-galactosyltransferase homology region (residues 114 to 378 of SEQ ID NO:2), and significant domains or regions in this family. Such fusion proteins would be expected to have a biological functional profile that is the same or similar to polypeptides of the present invention or other known galactosyltransferase family proteins (e.g. HSGALT3, HSGALT4, and *Brainiac*), depending on the fusion constructed. Moreover, such fusion proteins may exhibit other properties as disclosed herein.

Znssp6 polypeptides or fragments thereof may also be prepared through chemical synthesis. Znssp6 polypeptides may be monomers or multimers; glycosylated or non-glycosylated; pegylated or non-pegylated; and may or may not include an initial methionine amino acid residue.

Znssp6 polypeptides of the present invention can also be synthesized by exclusive solid phase synthesis, partial solid phase methods, fragment condensation or classical solution synthesis. The polypeptides are preferably prepared by solid phase peptide synthesis, for example as described by Merrifield, J. Am. Chem. Soc. 85:2149, 1963. The synthesis is carried out with amino acids that are protected at the alpha-amino terminus. Trifunctional amino acids with labile side-chains are also protected with suitable groups to prevent undesired chemical reactions from occurring during the assembly of the polypeptides. The alpha-amino protecting group is selectively removed to allow subsequent reaction to take place at the amino-terminus. The conditions for the removal of the alpha-amino protecting group do not remove the side-chain protecting groups.

The alpha-amino protecting groups are those known to be useful in the art of stepwise polypeptide synthesis. Included are acyl type protecting groups (e.g., formyl, trifluoroacetyl, acetyl), aryl type protecting groups (e.g., biotinyl), aromatic urethane type protecting groups [e.g., benzyloxycarbonyl (Cbz), substituted

benzyloxycarbonyl and 9-fluorenylmethyloxy-carbonyl (Fmoc)], aliphatic urethane protecting groups [e.g., t-butyloxycarbonyl (tBoc), isopropylloxycarbonyl, cyclohexylloxycarbonyl] and alkyl type protecting groups (e.g., benzyl, triphenylmethyl). The preferred protecting groups are tBoc and Fmoc, thus the peptides are said to be synthesized by tBoc and Fmoc chemistry, respectively.

The side-chain protecting groups selected must remain intact during coupling and not be removed during the deprotection of the amino-terminus protecting group or during coupling conditions. The side-chain protecting groups must also be removable upon the completion of synthesis using reaction conditions that will not alter the finished polypeptide. In tBoc chemistry, the side-chain protecting groups for trifunctional amino acids are mostly benzyl based. In Fmoc chemistry, they are mostly tert-butyl or trityl based.

In tBoc chemistry, the preferred side-chain protecting groups are tosyl for arginine, cyclohexyl for aspartic acid, 4-methylbenzyl (and acetamidomethyl) for cysteine, benzyl for glutamic acid, serine and threonine, benzyloxymethyl (and dinitrophenyl) for histidine, 2-Cl-benzyloxycarbonyl for lysine, formyl for tryptophan and 2-bromobenzyl for tyrosine. In Fmoc chemistry, the preferred side-chain protecting groups are 2,2,5,7,8-pentamethylchroman-6-sulfonyl (Pmc) or 2,2,4,6,7-pentamethyldihydrobenzofuran-5-sulfonyl (Pbf) for arginine, trityl for asparagine, cysteine, glutamine and histidine, tert-butyl for aspartic acid, glutamic acid, serine, threonine and tyrosine, tBoc for lysine and tryptophan.

For the synthesis of phosphopeptides, either direct or post-assembly incorporation of the phosphate group is used. In the direct incorporation strategy, the phosphate group on serine, threonine or tyrosine may be protected by methyl, benzyl, or tert-butyl in Fmoc chemistry or by methyl, benzyl or phenyl in tBoc chemistry. Direct incorporation of phosphotyrosine without phosphate protection can also be used in Fmoc chemistry. In the post-assembly incorporation strategy, the unprotected hydroxyl groups of serine, threonine or tyrosine are derivatized on solid phase with di-tert-butyl-, dibenzyl- or dimethyl-N,N'-diisopropylphosphoramidite and then oxidized by tert-butylhydroperoxide.

Solid phase synthesis is usually carried out from the carboxyl-terminus by coupling the alpha-amino protected (side-chain protected) amino acid to a suitable solid support. An ester linkage is formed when the attachment is made to a chloromethyl, chlorotriyl or hydroxymethyl resin, and the resulting polypeptide will have a free carboxyl group at the C-terminus. Alternatively, when an amide resin such as benzhydrylamine or p-methylbenzhydrylamine resin (for tBoc chemistry) and Rink amide or PAL resin (for Fmoc chemistry) are used, an amide bond is formed and the resulting polypeptide will have a carboxamide group at the C-terminus. These resins, whether polystyrene- or polyamide-based or polyethyleneglycol-grafted, with or without a handle or linker, with or without the first amino acid attached, are commercially available, and their preparations have been described by Stewart et al., "Solid Phase Peptide Synthesis" (2nd Edition), (Pierce Chemical Co., Rockford, IL, 1984) and Bayer & Rapp Chem. Pept. Prot. 3:3 (1986); and Atherton et al., Solid Phase Peptide Synthesis: A Practical Approach, IRL Press, Oxford, 1989.

The C-terminal amino acid, protected at the side chain if necessary, and at the alpha-amino group, is attached to a hydroxymethyl resin using various activating agents including dicyclohexylcarbodiimide (DCC), N,N'-diisopropylcarbodiimide (DIPCDI) and carbonyldiimidazole (CDI). It can be attached to chloromethyl or chlorotriyl resin directly in its cesium tetramethylammonium salt form or in the presence of triethylamine (TEA) or diisopropylethylamine (DIEA). First amino acid attachment to an amide resin is the same as amide bond formation during coupling reactions.

Following the attachment to the resin support, the alpha-amino protecting group is removed using various reagents depending on the protecting chemistry (e.g., tBoc, Fmoc). The extent of Fmoc removal can be monitored at 300-320 nm or by a conductivity cell. After removal of the alpha-amino protecting group, the remaining protected amino acids are coupled stepwise in the required order to obtain the desired sequence.

Various activating agents can be used for the coupling reactions including DCC, DIPCDI, 2-chloro-1,3-dimethylimidium hexafluorophosphate (CIP), benzotriazol-1-yl-oxy-tris-(dimethylamino)-phosphonium hexafluoro-phosphate (BOP)

and its pyrrolidine analog (PyBOP), bromo-tris-pyrrolidino-phosphonium hexafluorophosphate (PyBroP), O-(benzotriazol-1-yl)-1,1,3,3-tetramethyl-uronium hexafluorophosphate (HBTU) and its tetrafluoroborate analog (TBTU) or its pyrrolidine analog (HBPYU), O-(7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyl-uronium
 5 hexafluorophosphate (HATU) and its tetrafluoroborate analog (TATU) or its pyrrolidine analog (HAPYU). The most common catalytic additives used in coupling reactions include 4-dimethylaminopyridine (DMAP), 3-hydroxy-3,4-dihydro-4-oxo-1,2,3-benzotriazine (HODhbt), N-hydroxybenzotriazole (HOBt) and 1-hydroxy-7-azabenzotriazole (HOAt). Each protected amino acid is used in excess (>2.0
 10 equivalents), and the couplings are usually carried out in N-methylpyrrolidone (NMP) or in DMF, CH₂Cl₂ or mixtures thereof. The extent of completion of the coupling reaction can be monitored at each stage, e.g., by the ninhydrin reaction as described by Kaiser et al., Anal. Biochem. 34:595, 1970. In cases where incomplete coupling is found, the coupling reaction is extended and repeated and may have chaotropic salts
 15 added. The coupling reactions can be performed automatically with commercially available instruments such as ABI model 430A, 431A and 433A peptide synthesizers.

After the entire assembly of the desired peptide, the peptide-resin is cleaved with a reagent with proper scavengers. The Fmoc peptides are usually cleaved and deprotected by TFA with scavengers (e.g., H₂O, ethanedithiol, phenol and
 20 thioanisole). The tBoc peptides are usually cleaved and deprotected with liquid HF for 1-2 hours at -5 to 0° C, which cleaves the polypeptide from the resin and removes most of the side-chain protecting groups. Scavengers such as anisole, dimethylsulfide and p-thiocresol are usually used with the liquid HF to prevent cations formed during the cleavage from alkylating and acylating the amino acid residues present in the
 25 polypeptide. The formyl group of tryptophan and the dinitrophenyl group of histidine need to be removed, respectively by piperidine and thiophenyl in DMF prior to the HF cleavage. The acetamidomethyl group of cysteine can be removed by mercury(II)acetate and alternatively by iodine, thallium(III)trifluoroacetate or silver tetrafluoroborate which simultaneously oxidize cysteine to cystine. Other strong acids
 30 used for tBoc peptide cleavage and deprotection include trifluoromethanesulfonic acid (TFMSA) and trimethylsilyltrifluoroacetate (TMSOTf).

The activity of molecules of the present invention can be measured using a variety of assays that measure, for example, cell-cell interactions, glycolipid and glycoprotein biosynthesis, development, and other biological functions associated with galactosyltransferase family members. Of particular interest are changes in the transfer of galactosyl molecules in glycoprotein synthesis and in cell-cell interactions in brain, kidney, and testis tissue and cell lines derived from these tissues. Such assays are well known in the art. For a general reference, see Kolbinger, F. et al., J. Biol. Chem. 273: 433-440, 1998; Amado, M. et al., J. Biol. Chem. 273:12770-12778, 1998; Hennet, T. et al., J. Biol. Chem. 273:58-65, 1998; and Ram B.P., and Munjal, D.D., CRC Crit. Rev. Biochem. 17:257-311, 1985. Specific assays include, but are not limited to bioassays measuring cell migration, contact inhibition, tissue interactions, neuronal specificity, fertilization, embryonic cell adhesions, limb bud morphogenesis, mesenchyme development, immune recognition, growth control, tumor metastasis and suppression, and glycoprotein and glycolipid biosynthesis.

Additional activities likely associated with the polypeptides of the present invention include proliferation of cells of the brain, kidney, and testis directly or indirectly through other growth factors; action as a chemotaxic factor; and as a factor for expanding neuronal and epithelial stem cell and precursor populations.

Another assay of interest measures or detects changes in proliferation, differentiation, and development. Proliferation can be measured using cultured primary brain, kidney, and testis cells, *ex plant* tissues, or *in vivo* by administering molecules of the claimed invention to the appropriate cells, tissues, or animal models. Generally, proliferative effects are observed as an increase in cell number and therefore, may include inhibition of apoptosis, as well as mitogenesis. Likewise, a decrease in cell number and cell migration could be analyzed. Assays measuring cell proliferation are well known in the art. For example, assays measuring proliferation include such assays as chemosensitivity to neutral red dye (Cavanaugh et al., Investigational New Drugs 8:347-354, 1990,), incorporation of radiolabelled nucleotides (Cook et al., Analytical Biochem. 179:1-7, 1989,), incorporation of 5-bromo-2'-deoxyuridine (BrdU) in the DNA of proliferating cells (Porstmann et al., J. Immunol. Methods 82:169-179, 1985), and use of tetrazolium salts (Mosmann, J. Immunol. Methods 65:55-63, 1983; Alley et

al., Cancer Res. 48:589-601, 1988; Marshall et al., Growth Reg. 5:69-84, 1995; and Scudiero et al., Cancer Res. 48:4827-4833, 1988).

Proliferation of bone marrow and peripheral blood lymphocyte cells can be assayed by harvesting these cells from mice, suspending the mononuclear cells in a base medium, and measuring proliferation in the presence of znssp6 protein. Similarly, clonogenic assays can be performed.

To determine if znssp6 is a chemotractant *in vivo*, znssp6 can be given by intradermal or intraperitoneal injection. Characterization of the accumulated leukocytes at the site of injection can be determined using lineage specific cell surface markers and fluorescence immunocytometry or by immunohistochemistry (Jose, J. Exp. Med. 179:881-87, 1994). Release of specific leukocyte cell populations from bone marrow into peripheral blood can also be measured after znssp6 injection.

Differentiation is a progressive and dynamic process, beginning with pluripotent stem cells and ending with terminally differentiated cells. Pluripotent stem cells that can regenerate without commitment to a lineage express a set of differentiation markers that are lost when commitment to a cell lineage is made. Progenitor cells express a set of differentiation markers that may or may not continue to be expressed as the cells progress down the cell lineage pathway toward maturation. Differentiation markers that are expressed exclusively by mature cells are usually functional properties such as cell products, enzymes to produce cell products and receptors and receptor-like complementary molecules. The stage of a cell population's differentiation is monitored by identification of markers present in the cell population. For example, myocytes, osteoblasts, adipocytes, chondrocytes, fibroblasts and reticular cells are believed to originate from a common mesenchymal stem cell (Owen et al., Ciba Fdn. Symp. 136:42-46, 1988). Markers for mesenchymal stem cells have not been well identified (Owen et al., J. of Cell Sci. 87:731-738, 1987), so identification is usually made at the progenitor and mature cell stages.

There is evidence to suggest that factors that stimulate specific cell types down a pathway towards terminal differentiation or dedifferentiation affect the entire cell population originating from a common precursor or stem cell. Thus, znssp6 polypeptides may stimulate inhibition or proliferation of endocrine and exocrine cells

of the brain, kidney and testis, as well as, cells associated with the spinal cord, colon, prostate, stomach, ovary, pancreas, pituitary gland, adrenal gland, salivary gland, mammary gland, liver, small intestine, spleen, thymus, peripheral leukocyte, lymph node, bone marrow, lung, trachea, placenta, fetal spleen and fetal lung.

5 The znssp6 molecules of the present invention may, while stimulating proliferation or differentiation of brain, kidney and testis cells, inhibit proliferation or differentiation of other tissues, by virtue of their effect on common precursor/stem cells. The novel polypeptides of the present invention are useful to study neural and epithelial stem cells and brain, kidney and testis progenitor cells, both *in vivo* and *ex vivo*.

10 Assays measuring differentiation include, for example, measuring cell-surface markers associated with stage-specific expression of a tissue, enzymatic activity, functional activity or morphological changes (Watt, FASEB, 5:281-284, 1991; Francis, Differentiation 57:63-75, 1994; Raes, Adv. Anim. Cell Biol. Technol. Bioprocesses, 161-171, 1989).

15 The znssp6 polypeptides of the present invention can be used to study brain, kidney, and testis proliferation or differentiation. Such methods of the present invention generally comprise incubating cells derived from these tissues in the presence and absence of znssp6 polypeptide, monoclonal antibody, agonist or antagonist thereof and observing changes in cell proliferation or differentiation.

20 Proteins, including alternatively spliced peptides, and fragments, of the present invention are useful for cell-cell interactions, neuronal specificity, fertilization, morphogenesis, development, immune recognition, growth control, tumor suppression, and glycoprotein and glycolipid biosynthesis. Znssp6 molecules, variants, and fragments can be applied in isolation, or in conjunction with other molecules (growth factors, cytokines, etc.) in brain, kidney, and testis. Alternative splicing of znssp6 may be cell-type specific and confer activity to specific tissues.

25 Cell lines of brain, kidney, and testis are available from commercial manufacturers, such as the American Type Culture Collection. One skilled in the art would know how to order and establish such cell lines, and perform assays as described herein. Exemplary cell lines from ATCC include ATCC# CRL-7773, a brain

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ganglioneuroblastoma; ATCC# HTB-148, a brain neuoglioma; ATCC# CRL-7540, a kidney hypernephroma; ATCC# CRL-7192, a kidney carcinoma; and ATCC# CRL-7800, a testis seminoma, (ATCC, Manassas, VA).

Other assays to measure the effects of znssp6 include proliferation
5 assays (i.e., of brain, kidney, and testis) by testing tissue and cells from healthy volunteers with znssp6 protein, or a znssp6-free negative control for the ability of the tissue and cells to proliferate.

Proteins of the present invention are useful for delivery of therapeutic agents such as, but not limited to, radionuclides, chemotherapy agents, and small
10 molecules. The effects of znssp6 can be measured *in vitro* using cultured cells, *ex vivo* on tissue slices, or *in vivo* by administering molecules of the claimed invention to the appropriate animal model. For instance, znssp6 transfected (or co-transfected) expression host cells may be embedded in an alginate environment and injected (implanted) into recipient animals. Alginate-poly-L-lysine microencapsulation,
15 permselective membrane encapsulation and diffusion chambers have been described as a means to entrap transfected mammalian cells or primary mammalian cells. These types of non-immunogenic "encapsulations" or microenvironments permit the transfer of nutrients into the microenvironment, and also permit the diffusion of proteins and other macromolecules secreted or released by the captured cells across the
20 environmental barrier to the recipient animal. Most importantly, the capsules or microenvironments mask and shield the foreign, embedded cells from the recipient animal's immune response. Such microenvironments can extend the life of the injected cells from a few hours or days (naked cells) to several weeks (embedded cells).

Alginate threads provide a simple and quick means for generating
25 embedded cells. The materials needed to generate the alginate threads are readily available and relatively inexpensive. Once made, the alginate threads are relatively strong and durable, both *in vitro* and, based on data obtained using the threads, *in vivo*. The alginate threads are easily manipulable and the methodology is scalable for preparation of numerous threads. In an exemplary procedure, 3% alginate is prepared
30 in sterile H₂O, and sterile filtered. Just prior to preparation of alginate threads, the alginate solution is again filtered. An approximately 50% cell suspension (containing

about 5×10^5 to about 5×10^7 cells/ml) is mixed with the 3% alginate solution. One ml of the alginate/cell suspension is extruded into a 100 mM sterile filtered CaCl_2 solution over a time period of ~15 min, forming a "thread". The extruded thread is then transferred into a solution of 50 mM CaCl_2 , and then into a solution of 25 mM CaCl_2 .

- 5 The thread is then rinsed with deionized water before coating the thread by incubating in a 0.01% solution of poly-L-lysine. Finally, the thread is rinsed with Lactated Ringer's Solution and drawn from solution into a syringe barrel (without needle attached). A large bore needle is then attached to the syringe, and the thread is intraperitoneally injected into a recipient in a minimal volume of the Lactated Ringer's
- 10 Solution.

An alternative *in vivo* approach for assaying proteins of the present invention involves viral delivery systems. Exemplary viruses for this purpose include adenovirus, herpesvirus, lentivirus, vaccinia virus and adeno-associated virus (AAV). Adenovirus, a double-stranded DNA virus, is currently the best studied gene transfer

- 15 vector for delivery of heterologous nucleic acid (for a review, see T.C. Becker et al., Meth. Cell Biol. 43:161-89, 1994; and J.T. Douglas and D.T. Curiel, Science & Medicine 4:44-53, 1997). The adenovirus system offers several advantages: adenovirus can (i) accommodate relatively large DNA inserts; (ii) be grown to high-titer; (iii) infect a broad range of mammalian cell types; and (iv) be used with a large
- 20 number of available vectors containing different promoters. Also, because adenoviruses are stable in the bloodstream, they can be administered by intravenous injection.

By deleting portions of the adenovirus genome, larger inserts (up to 7 kb) of heterologous DNA can be accommodated. These inserts can be incorporated

- 25 into the viral DNA by direct ligation or by homologous recombination with a co-transfected plasmid. In an exemplary system, the essential E1 gene has been deleted from the viral vector, and the virus will not replicate unless the E1 gene is provided by the host cell (the human 293 cell line is exemplary). When intravenously administered to intact animals, adenovirus primarily targets the liver. If the adenoviral delivery
- 30 system has an E1 gene deletion, the virus cannot replicate in the host cells. However,

the host's tissue (e.g., liver) will express and process (and, if a secretory signal sequence is present, secrete) the heterologous protein. Secreted proteins will enter the circulation in the highly vascularized liver, and effects on the infected animal can be determined.

5 The adenovirus system can also be used for protein production *in vitro*. By culturing adenovirus-infected non-293 cells under conditions where the cells are not rapidly dividing, the cells can produce proteins for extended periods of time. For instance, BHK cells are grown to confluence in cell factories, then exposed to the adenoviral vector encoding the secreted protein of interest. The cells are then grown
10 under serum-free conditions, which allows infected cells to survive for several weeks without significant cell division. Alternatively, adenovirus vector infected 293S cells can be grown in suspension culture at relatively high cell density to produce significant amounts of protein (see Garnier et al., Cytotechnol. 15:145-55, 1994). With either protocol, an expressed, secreted heterologous protein can be repeatedly isolated from
15 the cell culture supernatant. Within the infected 293S cell production protocol, non-secreted proteins may also be effectively obtained.

In view of the tissue distribution (i.e., brain, kidney, and testis and various other tissues) observed for znssp6, agonists (including the natural anti-complementary molecule) and antagonists have enormous potential for both *in vitro*
20 and *in vivo* applications. Compounds identified as znssp6 agonists are useful for studying galactosylation of cell surface antigens as well as cell-cell interactions *in vitro* and *in vivo*. For example, znssp6 and agonist compounds are useful as components of defined cell culture media, and may be used alone or in combination with other cytokines and hormones to replace serum that is commonly used in cell culture.
25 Agonists are thus useful in specifically promoting the growth and/or development of brain, kidney, and testis in culture. Alternatively, znssp6 polypeptides and znssp6 agonist polypeptides are useful as a research reagent, particularly for the growth and expansion of neural and epithelial cells. Znssp6 polypeptides are added to tissue culture media for these cell types.

30 Additionally, molecules of the present invention can be used *in vitro* to modify glycoproteins. Expression of proteins which will be used as therapeutics can

result in aberrant glycosylation. Znssp6 molecules can be added *in vitro* to production or reagent grade proteins to modify the improper galactosylation of proteins.

Antagonists of znssp6 molecules are also useful as research reagents for characterizing sites of interactions between member of complement/anti-complement
5 pairs as well as site of galactosyltransferase catalysis.

Inhibitors of znssp6 activity (znssp6 antagonists) include anti-znssp6 antibodies and soluble znssp6 polypeptides, as well as other peptidic and non-peptidic agents (including ribozymes).

The invention also provides antagonists, which either bind to znssp6
10 polypeptides or, alternatively, to a anti-complementary molecule to which znssp6 polypeptides bind, thereby inhibiting or eliminating the function of znssp6. Such znssp6 antagonists would include antibodies; polypeptides which bind either to the znssp6 polypeptide or to its anti-complementary molecule or natural or synthetic analogs of znssp6 anti-complementary molecule which retain the ability to bind the
15 anti-complementary molecule but do not result in glycoprotein or glycolipid synthesis or cell-cell interactions. Such analogs could be peptides or peptide-like compounds. Natural or synthetic small molecules which bind to znssp6 polypeptides and prevent glycoprotein or glycolipid synthesis or cell-cell interactions are also contemplated as antagonists. Also contemplated are soluble znssp6 polypeptides. As such, znssp6
20 antagonists would be useful as therapeutics for treating certain disorders where blocking glycosylation or binding of the znssp6-anti-complementary molecule would be beneficial.

Znssp6 polypeptides may be used within diagnostic systems to detect the presence of znssp6 anti-complementary molecule polypeptides. Antibodies or other
25 agents that specifically bind to znssp6 or its anti-complementary molecule may also be used to detect the presence of circulating znssp6 or anti-complementary molecule polypeptides. Such detection methods are well known in the art and include, for example, enzyme-linked immunosorbent assay (ELISA) and radioimmunoassay. Immunohistochemically labeled znssp6 antibodies can be used to detect znssp6 and/or
30 znssp6 anti-complementary molecule in tissue samples. znssp6 levels can also be monitored by such methods as RT-PCR, where znssp6 mRNA can be detected and

quantified. The information derived from such detection methods would provide insight into the significance of znssp6 polypeptides in various diseases, and as such would serve as diagnostic tools for diseases for which altered levels of znssp6 are significant. Altered levels of znssp6 polypeptides may be indicative of pathological conditions including, for example, cancer, auto-immune diseases, digestive disorders and inflammatory disorders.

A "soluble protein" is a protein that is not bound to a cell membrane. Soluble znssp6 proteins are most commonly anti-complementary molecule-binding polypeptides that lack transmembrane and cytoplasmic domains. Soluble proteins can comprise additional amino acid residues, such as affinity tags that provide for purification of the polypeptide or provide sites for attachment of the polypeptide to a anti-complementary molecule, or immunoglobulin constant region sequences. Many cell-surface proteins have naturally occurring, soluble counterparts that are produced by proteolysis or translated from alternatively spliced mRNAs. Proteins are said to be substantially free of transmembrane and intracellular polypeptide segments when they lack sufficient portions of these segments to provide membrane anchoring or signal transduction, respectively.

Soluble forms of znssp6 polypeptides can be generated by removing the hydrophobic region between residues 26 and 49 of SEQ ID NO: 2. Soluble znssp6 polypeptides are useful in studying the effects of the present invention *in vivo* and *in vitro*. Such soluble molecules comprise the anti-complmentary moleculc binding domain comprising reisudes 114 to 370 of SEQ ID NO:2. Thus, soluble forms of znssp6 can also include polypeptides selected from the rest of the molecule excluding the hydrophobic region. Some exemplary forms of znssp6 polypeptides include: the polypeptide from residue 114 to 370 fo SEQ ID NO:2; the polypeptide from residue 114 to residue 378 of SEQ ID NO:2; the polypeptide from residue 50 to residue 370 of SEQ ID NO:2; and the polypeptide from residue 50 to residue 378 of SEQ ID NO:2.

Soluble forms of znssp6 polypeptides may act as antagonists to or agonists of znssp6 polypeptides, and would be useful to modulate the effects of znssp6 in brain, kidney and testis. Since polypeptides of this nature are not anchored to the membrane, they can act at sites distant from the tissues in which they are expressed.

Thus, the activity of the soluble form of znssp6 polypeptides can be more wide spread than its membrane-anchored counterpart. Both isoforms would be useful in studying the effects of the present invention *in vitro* an *in vivo*.

Znssp6 can also be used to identify inhibitors (antagonists) of its
5 activity. Test compounds are added to the assays disclosed herein to identify compounds that inhibit the activity of znssp6. In addition to those assays disclosed herein, samples can be tested for inhibition of znssp6 activity within a variety of assays designed to measure binding or the stimulation/inhibition of znssp6-dependent cellular responses. For example, znssp6-responsive cell lines can be transfected with a reporter
10 gene construct that is responsive to a znssp6-stimulated cellular pathway. Reporter gene constructs of this type are known in the art, and will generally comprise a znssp6-DNA response element operably linked to a gene encoding an assayable protein, such as luciferase. DNA response elements can include, but are not limited to, cyclic AMP response elements (CRE), hormone response elements (HRE) insulin response element
15 (IRE) (Nasrin et al., Proc. Natl. Acad. Sci. USA 87:5273-7, 1990) and serum response elements (SRE) (Shaw et al. Cell 56: 563-72, 1989). Cyclic AMP response elements are reviewed in Roestler et al., J. Biol. Chem. 263 (19):9063-6; 1988 and Habener, Molec. Endocrinol. 4 (8):1087-94; 1990. Hormone response elements are reviewed in Beato, Cell 56:335-44; 1989. Candidate compounds, solutions, mixtures or extracts are
20 tested for the ability to inhibit the activity of znssp6 on the target cells as evidenced by a decrease in znssp6 stimulation of reporter gene expression. Assays of this type will detect compounds that directly block znssp6 binding to cell-surface anti-complementary molecule, as well as compounds that block processes in the cellular pathway subsequent to complementary molecule/anti-complementary molecule
25 binding. In the alternative, compounds or other samples can be tested for direct blocking of znssp6 binding to its anti-complementary molecule using znssp6 tagged with a detectable label (e.g., ¹²⁵I, biotin, horseradish peroxidase, FITC, and the like). Within assays of this type, the ability of a test sample to inhibit the binding of labeled znssp6 to the anti-complementary molecule is indicative of inhibitory activity, which
30 can be confirmed through secondary assays. Anti-complementary molecules used

within binding assays may be cellular or isolated, immobilized, or receptor-like complementary molecules.

Assays measuring the inhibition of galactosyltransferase activity in glycoprotein synthesis are listed in Ram, B.P., (*ibid*).

5 Also, znssp6 polypeptides, agonists or antagonists thereof may be therapeutically useful for promoting wound healing, for example, in brain, kidney and testis tissues. To verify the presence of this capability in znssp6 polypeptides, agonists or antagonists of the present invention, such znssp6 polypeptides, agonists or antagonists are evaluated with respect to their ability to facilitate wound healing
10 according to procedures known in the art. If desired, znssp6 polypeptide performance in this regard can be compared to growth factors, such as EGF, NGF, TGF- α , TGF- β , insulin, IGF-I, IGF-II, fibroblast growth factor (FGF) and the like. In addition, znssp6 polypeptides or agonists or antagonists thereof may be evaluated in combination with one or more growth factors to identify synergistic effects.

15 A znssp6 polypeptide can be expressed as a fusion with an immunoglobulin heavy chain constant region, typically an F_C fragment, which contains two constant region domains and lacks the variable region. Methods for preparing such fusions are disclosed in U.S. Patents Nos. 5,155,027 and 5,567,584. Such fusions are typically secreted as multimeric molecules wherein the F_C portions are disulfide bonded
20 to each other and two non-Ig polypeptides are arrayed in closed proximity to each other. Fusions of this type can be used to evaluate specific donor/acceptor molecules, affinity purify ligands, or use as an *in vitro* assay tool. This fusion can also be used to determine the homodimerization potential for znssp6. For use in assays, the chimeras are bound to a support via the F_C region and used in an ELISA format.

25 A znssp6 polypeptide can also be used for purification of its anti-complementary molecule. The polypeptide is immobilized on a solid support, such as beads of agarose, cross-linked agarose, glass, cellulosic resins, silica-based resins, polystyrene, cross-linked polyacrylamide, or like materials that are stable under the conditions of use. Methods for linking polypeptides to solid supports are known in the
30 art, and include amine chemistry, cyanogen bromide activation, N-hydroxysuccinimide activation, epoxide activation, sulfhydryl activation, and hydrazide activation. The

resulting medium will generally be configured in the form of a column or chip, and fluids containing the anti-complementary molecule are passed through the column or chip one or more times to allow the znssp6 polypeptide to bind to the anti-complementary molecule. The anti-complementary molecule is then eluted using
 5 changes in salt concentration, chaotropic agents (guanidine HCl), or pH to disrupt znssp6- anti-complementary molecule binding.

An assay system that uses a ligand-binding receptor (or an antibody, one member of a complement/ anti-complement pair) or a binding fragment thereof, and a commercially available biosensor instrument (BIAcore, Pharmacia Biosensor,
 10 Piscataway, NJ) may be advantageously employed. Such receptor, antibody, member of a complement/anti-complement pair or fragment is immobilized onto the surface of a receptor chip. Use of this instrument is disclosed by Karlsson, J. Immunol. Methods 145:229-40, 1991 and Cunningham and Wells, J. Mol. Biol. 234:554-63, 1993. A receptor, antibody, member or fragment is covalently attached, using amine or
 15 sulfhydryl chemistry, to dextran fibers that are attached to gold film within the flow cell. A test sample is passed through the cell. If a ligand, epitope, or opposite member of the complement/anti-complement pair is present in the sample, it will bind to the immobilized receptor, antibody or member, respectively, causing a change in the refractive index of the medium, which is detected as a change in surface plasmon
 20 resonance of the gold film. This system allows the determination of on- and off-rates, from which binding affinity can be calculated, and assessment of stoichiometry of binding.

Znssp6 polypeptides can also be used within other assay systems known in the art. Such systems include Scatchard analysis for determination of binding
 25 affinity (see Scatchard, Ann. NY Acad. Sci. 51: 660-72, 1949) and calorimetric assays (Cunningham et al., Science 253:545-48, 1991; Cunningham et al., Science 245:821-25, 1991).

Within the polypeptides of the present invention are polypeptides that comprise an epitope-bearing portion of a protein as shown in SEQ ID NO:2. An
 30 "epitope" is a region of a protein to which an antibody can bind. See, for example, Geysen et al., Proc. Natl. Acad. Sci. USA 81:3998-4002, 1984. Epitopes can be linear

or conformational, the latter being composed of discontinuous regions of the protein that form an epitope upon folding of the protein. Linear epitopes are generally at least 6 amino acid residues in length. Relatively short synthetic peptides that mimic part of a protein sequence are routinely capable of eliciting an antiserum that reacts with the partially mimicked protein. See, Sutcliffe et al., Science 219:660-666, 1983. Antibodies that recognize short, linear epitopes are particularly useful in analytic and diagnostic applications that employ denatured protein, such as Western blotting (Tobin, Proc. Natl. Acad. Sci. USA 76:4350-4356, 1979), or in the analysis of fixed cells or tissue samples. Antibodies to linear epitopes are also useful for detecting fragments of znssp6, such as might occur in body fluids or cell culture media.

Antigenic, epitope-bearing polypeptides of the present invention are useful for raising antibodies, including monoclonal antibodies, that specifically bind to a znssp6 protein. The znssp6 polypeptide or a fragment thereof serves as an antigen (immunogen) to inoculate an animal and elicit an immune response. One of skill in the art would recognize that antigenic, epitope-bearing polypeptides contain a sequence of at least six, or at least nine, or from 15 to about 30 contiguous amino acid residues of a znssp6 protein (e.g., SEQ ID NO:2). Polypeptides comprising a larger portion of a znssp6 protein, i.e. from 30 to 100 residues up to the entire sequence, are included. Antigens or immunogenic epitopes can also include attached tags, adjuvants and carriers, as described herein. Suitable antigens include the znssp6 polypeptides encoded by SEQ ID NO:2 from amino acid number 1 to amino acid number 378, or a contiguous 9 to 378 amino acid fragment thereof. Such regions include the catalytic domain, the stem domain, or the hydrophobic domain of znssp6 and fragments thereof. Polypeptides in this regard include those comprising residues 1 to 25 of SEQ ID NO:2; residues 26 to 49 of SEQ ID NO:2; residues 50 to 113 of SEQ ID NO:2; residues 114 to 378 of SEQ ID NO:2; and residues 371 to 378 of SEQ ID NO:2.

The present invention also provides polypeptide fragments or peptides comprising an epitope-bearing portion of a znssp6 polypeptide described herein. Such fragments or peptides may comprise an "immunogenic epitope," which is a part of a protein that elicits an antibody response when the entire protein is used as an

immunogen. Immunogenic epitope-bearing peptides can be identified using standard methods (see, for example, Geysen *et al.*, Proc. Nat'l Acad. Sci. USA 81:3998 (1983)).

In contrast, polypeptide fragments or peptides may comprise an "antigenic epitope," which is a region of a protein molecule to which an antibody can specifically bind. Certain epitopes consist of a linear or contiguous stretch of amino acids, and the antigenicity of such an epitope is not disrupted by denaturing agents. It is known in the art that relatively short synthetic peptides that can mimic epitopes of a protein can be used to stimulate the production of antibodies against the protein (see, for example, Sutcliffe *et al.*, Science 219:660 (1983)). Accordingly, antigenic epitope-bearing peptides and polypeptides of the present invention are useful to raise antibodies that bind with the polypeptides described herein.

Antigenic epitope-bearing peptides and polypeptides contain at least four to ten amino acids, or at least ten to fifteen amino acids, or 15 to 30 amino acids of SEQ ID NO:2. Such epitope-bearing peptides and polypeptides can be produced by fragmenting an znssp6 polypeptide, or by chemical peptide synthesis, as described herein. Moreover, epitopes can be selected by phage display of random peptide libraries (see, for example, Lane and Stephen, Curr. Opin. Immunol. 5:268 (1993), and Cortese *et al.*, Curr. Opin. Biotechnol. 7:616 (1996)). Standard methods for identifying epitopes and producing antibodies from small peptides that comprise an epitope are described, for example, by Mole, "Epitope Mapping," in *Methods in Molecular Biology*, Vol. 10, Manson (ed.), pages 105-116 (The Humana Press, Inc. 1992), Price, "Production and Characterization of Synthetic Peptide-Derived Antibodies," in *Monoclonal Antibodies: Production, Engineering, and Clinical Application*, Ritter and Ladyman (eds.), pages 60-84 (Cambridge University Press 1995), and Coligan *et al.* (eds.), *Current Protocols in Immunology*, pages 9.3.1 - 9.3.5 and pages 9.4.1 - 9.4.11 (John Wiley & Sons 1997).

As an illustration, potential antigenic sites in znssp6 polypeptides (SEQ ID NO:2) were identified using the Jameson-Wolf method, Jameson and Wolf, *CABIOS* 4:181, (1988), as implemented by the PROTEAN program (version 3.14) of LASERGENE (DNASTAR; Madison, WI). Default parameters were used in this analysis.

Suitable antigens of znssp6 polypeptides include residue 10 to residue 16 of SEQ ID NO:2; residue 52 to residue 61 of SEQ ID NO:2; residue 52 to residue 78 of SEQ ID NO:2; residue 69 to residue 78 of SEQ ID NO:2; residue 89 to residue 94 of SEQ ID NO:2; residue 89 to residue 117 of SEQ ID NO:2; residue 111 to residue 117 of SEQ ID NO:2; residue 126 to residue 134 of SEQ ID NO:2; residue 126 to residue 151 of SEQ ID NO:2; residue 143 to residue 151 of SEQ ID NO:2; residue 215 to residue 220 of SEQ ID NO:2; residue 215 to residue 239 of SEQ ID NO:2; residue 223 to residue 239 of SEQ ID NO:2; residue 223 to residue 257 of SEQ ID NO:2; residue 251 to residue 257 of SEQ ID NO:2; and residue 332 to residue 337 of SEQ ID NO:2; or a portion thereof which contains a 4 to 10 amino acid segment. Hydrophilic peptides, such as those predicted by one of skill in the art from a hydrophobicity plot are also immunogenic. Znssp6 hydrophilic peptides include peptides comprising amino acid sequences selected from the group consisting of: residue 6 to residue 20 of SEQ ID NO:2; residue 7 to residue 15 of SEQ ID NO:2; residue 52 to residue 82 of SEQ ID NO:2; residue 52 to residue 69 of SEQ ID NO:2; residue 54 to residue 62 of SEQ ID NO:2; residue 62 to residue 82 of SEQ ID NO:2; residue 69 to residue 79 of SEQ ID NO:2; residue 89 to residue 104 of SEQ ID NO:2; residue 97 to residue 104 of SEQ ID NO:2; residue 126 to residue 153 of SEQ ID NO:2; residue 129 to residue 134 of SEQ ID NO:2; residue 126 to residue 140 of SEQ ID NO:2; residue 135 to residue 140 of SEQ ID NO:2; residue 141 to residue 153 of SEQ ID NO:2; residue 172 to residue 181 of SEQ ID NO:2; residue 172 to residue 189 of SEQ ID NO:2; residue 175 to residue 181 of SEQ ID NO:2; residue 175 to residue 189 of SEQ ID NO:2; residue 213 to residue 218 of SEQ ID NO:2; residue 233 to residue 238 of SEQ ID NO:2; residue 249 to residue 261 of SEQ ID NO:2; residue 249 to residue 257 of SEQ ID NO:2; residue 251 to residue 257 of SEQ ID NO:2; residue 251 to residue 261 of SEQ ID NO:2; residue 266 to residue 272 of SEQ ID NO:2; residue 266 to residue 277 of SEQ ID NO:2; residue 328 to residue 336 of SEQ ID NO:2; and residue 373 to residue 378 of SEQ ID NO:2; or a portion thereof which contains a 4 to 10 amino acid segment. Additionally, antigens can be generated to portions of the polypeptide which are likely to be on the surface of the folded protein. These antigens include: residue 3 to residue 8 of SEQ ID NO:2; residue 3 to residue 15 of SEQ ID NO:2; residue 10 to residue 15 of SEQ ID

NO:2; residue 51 to residue 62 of SEQ ID NO:2; residue 51 to residue 77 of SEQ ID NO:2; residue 68 to residue 77 SEQ ID NO:2; residue 129 to residue 135 of SEQ ID NO:2; residue 173 to residue 178 of SEQ ID NO:2; residue 250 to residue 258 of SEQ ID NO:2; residue 250 to residue 274 SEQ ID NO:2; and residue 265 to residue 274 of
 5 SEQ ID NO:2; or a portion thereof which contains a 4 to 10 amino acid segment.

Antibodies generated from this immune response can be isolated and purified as described herein. Methods for preparing and isolating polyclonal and monoclonal antibodies are well known in the art. See, for example, Current Protocols in Immunology, Cooligan, et al. (eds.), National Institutes of Health, John Wiley and
 10 Sons, Inc., 1995; Sambrook et al., Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor, NY, 1989; and Hurrell, J. G. R., Ed., Monoclonal Hybridoma Antibodies: Techniques and Applications, CRC Press, Inc., Boca Raton, FL, 1982.

As would be evident to one of ordinary skill in the art, polyclonal
 15 antibodies can be generated from inoculating a variety of warm-blooded animals such as horses, cows, goats, sheep, dogs, chickens, rabbits, mice, and rats with a znssp6 polypeptide or a fragment thereof. The immunogenicity of a znssp6 polypeptide may be increased through the use of an adjuvant, such as alum (aluminum hydroxide) or Freund's adjuvant. Polypeptides useful for immunization also include fusion
 20 polypeptides, such as fusions of znssp6 or a portion thereof with an immunoglobulin polypeptide or with maltose binding protein. The polypeptide immunogen may be a full-length molecule or a portion thereof. If the polypeptide portion is "hapten-like", such portion may be advantageously joined or linked to a macromolecular carrier (such as keyhole limpet hemocyanin (KLH), bovine serum albumin (BSA) or tetanus toxoid)
 25 for immunization.

As used herein, the term "antibodies" includes polyclonal antibodies, affinity-purified polyclonal antibodies, monoclonal antibodies, and antigen-binding fragments, such as F(ab')₂ and Fab proteolytic fragments. Genetically engineered intact antibodies or fragments, such as chimeric antibodies, Fv fragments, single chain
 30 antibodies and the like, as well as synthetic antigen-binding peptides and polypeptides, are also included. Non-human antibodies may be humanized by grafting non-human

CDRs onto human framework and constant regions, or by incorporating the entire non-human variable domains (optionally "cloaking" them with a human-like surface by replacement of exposed residues, wherein the result is a "veneered" antibody). In some instances, humanized antibodies may retain non-human residues within the human
5 variable region framework domains to enhance proper binding characteristics. Through humanizing antibodies, biological half-life may be increased, and the potential for adverse immune reactions upon administration to humans is reduced.

Alternative techniques for generating or selecting antibodies useful herein include *in vitro* exposure of lymphocytes to znssp6 protein or peptide, and
10 selection of antibody display libraries in phage or similar vectors (for instance, through use of immobilized or labeled znssp6 protein or peptide). Genes encoding polypeptides having potential znssp6 polypeptide binding domains can be obtained by screening random peptide libraries displayed on phage (phage display) or on bacteria, such as *E. coli*. Nucleotide sequences encoding the polypeptides can be obtained in a number of
15 ways, such as through random mutagenesis and random polynucleotide synthesis. These random peptide display libraries can be used to screen for peptides which interact with a known target which can be a protein or polypeptide, such as a ligand or receptor, a biological or synthetic macromolecule, or organic or inorganic substances. Techniques for creating and screening such random peptide display libraries are known
20 in the art (Ladner et al., US Patent NO. 5,223,409; Ladner et al., US Patent NO. 4,946,778; Ladner et al., US Patent NO. 5,403,484 and Ladner et al., US Patent NO. 5,571,698) and random peptide display libraries and kits for screening such libraries are available commercially, for instance from Clontech (Palo Alto, CA), Invitrogen Inc. (San Diego, CA), New England Biolabs, Inc. (Beverly, MA) and Pharmacia LKB
25 Biotechnology Inc. (Piscataway, NJ). Random peptide display libraries can be screened using the znssp6 sequences disclosed herein to identify proteins which bind to znssp6. These "binding proteins" which interact with znssp6 polypeptides can be used for tagging cells; for isolating homolog polypeptides by affinity purification; they can be directly or indirectly conjugated to drugs, toxins, radionuclides and the like. These
30 binding proteins can also be used in analytical methods such as for screening expression libraries and neutralizing activity. The binding proteins can also be used

for diagnostic assays for determining circulating levels of polypeptides; for detecting or quantitating soluble polypeptides as marker of underlying pathology or disease. These binding proteins can also act as znssp6 "antagonists" to block znssp6 binding and signal transduction *in vitro* and *in vivo*. These anti-znssp6 binding proteins would be useful
 5 for mediating galactosyltransferase activity extracellularly, therefore, mediating cell-cell interactions, such as, for example, tumor formation and metastasis, proliferation and differentiation, as well as glycoprotein synthesis.

As used herein, the term "binding proteins" additionally includes antibodies to znssp6 polypeptides, the cognate anti-complementary molecule of znssp6
 10 polypeptides, proteins useful for purification of znssp6 polypeptides, and proteins associated with the catalytic domain (residues 114 to 378 of SEQ ID NO:2).

Antibodies are determined to be specifically binding if: 1) they exhibit a threshold level of binding activity, and/or 2) they do not significantly cross-react with related polypeptide molecules. First, antibodies herein specifically bind if they bind to
 15 a znssp6 polypeptide, peptide or epitope with a binding affinity (K_a) of 10^6 M^{-1} or greater, preferably 10^7 M^{-1} or greater, more preferably 10^8 M^{-1} or greater, and most preferably 10^9 M^{-1} or greater. The binding affinity of an antibody can be readily determined by one of ordinary skill in the art, for example, by Scatchard analysis (Scatchard, G., Ann. NY Acad. Sci. 51: 660-672, 1949).

20 Second, antibodies are determined to specifically bind if they do not significantly cross-react with related polypeptides. Antibodies do not significantly cross-react with related polypeptide molecules, for example, if they detect znssp6 but not known related polypeptides using a standard Western blot analysis (Ausubel et al., *ibid.*). Examples of known related polypeptides are orthologs, proteins from the same
 25 species that are members of a protein family (e.g. IL-16), znssp6 polypeptides, and non-human znssp6. Moreover, antibodies may be "screened against" known related polypeptides to isolate a population that specifically binds to the inventive polypeptides. For example, antibodies raised to znssp6 are adsorbed to related polypeptides adhered to insoluble matrix; antibodies specific to znssp6 will flow
 30 through the matrix under the proper buffer conditions. Such screening allows isolation

of polyclonal and monoclonal antibodies non-crossreactive to closely related polypeptides (Antibodies: A Laboratory Manual, Harlow and Lane (eds.), Cold Spring Harbor Laboratory Press, 1988; Current Protocols in Immunology, Cooligan, et al. (eds.), National Institutes of Health, John Wiley and Sons, Inc., 1995). Screening and isolation of specific antibodies is well known in the art. See, Fundamental Immunology, Paul (eds.), Raven Press, 1993; Getzoff et al., *Adv. in Immunol.* 43: 1-98, 1988; Monoclonal Antibodies: Principles and Practice, Goding, J.W. (eds.), Academic Press Ltd., 1996; Benjamin et al., Ann. Rev. Immunol. 2: 67-101, 1984.

A variety of assays known to those skilled in the art can be utilized to detect antibodies which specifically bind to znssp6 proteins or peptides. Exemplary assays are described in detail in Antibodies: A Laboratory Manual, Harlow and Lane (Eds.), Cold Spring Harbor Laboratory Press, 1988. Representative examples of such assays include: concurrent immunoelectrophoresis, radioimmunoassay, radioimmuno-precipitation, enzyme-linked immunosorbent assay (ELISA), dot blot or Western blot assay, inhibition or competition assay, and sandwich assay. In addition, antibodies can be screened for binding to wild-type versus mutant znssp6 protein or polypeptide.

Antibodies to znssp6 may be used for tagging or labeling cells that express znssp6; for isolating znssp6 by affinity purification; for diagnostic assays for determining circulating levels of znssp6 polypeptides; for detecting or quantitating soluble znssp6 as marker of underlying pathology or disease; in analytical methods employing FACS; for screening expression libraries; for generating anti-idiotypic antibodies; and as neutralizing antibodies or as antagonists to block znssp6 *in vitro* and *in vivo*. Suitable direct tags or labels include radionuclides, enzymes, substrates, cofactors, inhibitors, fluorescent markers, chemiluminescent markers, magnetic particles and the like; indirect tags or labels may feature use of biotin-avidin or other complement/anti-complement pairs as intermediates. Antibodies herein may also be directly or indirectly conjugated to drugs, toxins, radionuclides and the like, and these conjugates used for *in vivo* diagnostic or therapeutic applications. Moreover, antibodies to znssp6 or fragments thereof may be used *in vitro* to detect denatured znssp6 or fragments thereof in assays, for example, Western Blots or other assays known in the art.

The soluble znssp6 is useful in studying the distribution of its anti-complementary molecule in tissues or specific cell lineages, and to provide insight into complementary molecule-anti-complementary molecule biology. Using labeled znssp6, cells expressing the anti-complementary molecule are identified by fluorescence
5 immunocytometry or immunocytochemistry. Application may also be made of the specificity of UDP-glycosyltransferases for their substrates.

Antibodies can be made to soluble, znssp6 polypeptides which are His or FLAGTM tagged. Alternatively, such polypeptides form a fusion protein with Human Ig. In particular, antiserum containing polypeptide antibodies to His-tagged, or
10 FLAGTM-tagged soluble znssp6 can be used in analysis of tissue distribution of znssp6 by immunohistochemistry on human or primate tissue. These soluble znssp6 polypeptides can also be used to immunize mice in order to produce monoclonal antibodies to a soluble human znssp6 polypeptide. Monoclonal antibodies to a soluble human znssp6 polypeptide can also be used to mimic anti-complementary molecule
15 coupling, resulting in activation or inactivation of the complementary molecule-anti-complementary molecule pair. For instance, it has been demonstrated that cross-linking anti-soluble CD40 monoclonal antibodies provides a stimulatory signal to B cells that have been sub-optimally activated with anti-IgM or LPS, and results in proliferation and immunoglobulin production. These same monoclonal antibodies act as antagonists
20 when used in solution by blocking activation of the receptor. Monoclonal antibodies to znssp6 can be used to determine the distribution, regulation and biological interaction of the znssp6 and its anti-complementary molecule pair on specific cell lineages identified by tissue distribution studies.

Antibodies or polypeptides herein can also be directly or indirectly
25 conjugated to drugs, toxins, radionuclides and the like, and these conjugates used for *in vivo* diagnostic or therapeutic applications. For instance, polypeptides or antibodies of the present invention can be used to identify or treat tissues or organs that express a corresponding anti-complementary molecule. More specifically, znssp6 polypeptides or anti-znssp6 antibodies, or bioactive fragments or portions thereof, can be coupled to
30 detectable or cytotoxic molecules and delivered to a mammal having cells, tissues or organs that express the anti-complementary molecule.

Suitable detectable molecules may be directly or indirectly attached to the polypeptide or antibody, and include radionuclides, enzymes, substrates, cofactors, inhibitors, fluorescent markers, chemiluminescent markers, magnetic particles and the like. Suitable cytotoxic molecules may be directly or indirectly attached to the polypeptide or antibody, and include bacterial or plant toxins (for instance, diphtheria toxin, *Pseudomonas* exotoxin, ricin, abrin and the like), as well as therapeutic radionuclides, such as iodine-131, rhenium-188 or yttrium-90 (either directly attached to the polypeptide or antibody, or indirectly attached through means of a chelating moiety, for instance). Polypeptides or antibodies may also be conjugated to cytotoxic drugs, such as adriamycin. For indirect attachment of a detectable or cytotoxic molecule, the detectable or cytotoxic molecule can be conjugated with a member of a complementary/ anticomplementary pair, where the other member is bound to the polypeptide or antibody portion. For these purposes, biotin/streptavidin is an exemplary complementary/ anticomplementary pair.

In another embodiment, polypeptide-toxin fusion proteins or antibody-toxin fusion proteins can be used for targeted cell or tissue inhibition or ablation (for instance, to treat cancer cells or tissues). Alternatively, if the polypeptide has multiple functional domains (i.e., an activation domain or a ligand binding domain, plus a targeting domain), a fusion protein including only the targeting domain may be suitable for directing a detectable molecule, a cytotoxic molecule or a complementary molecule to a cell or tissue type of interest. In instances where the domain only fusion protein includes a complementary molecule, the anti-complementary molecule can be conjugated to a detectable or cytotoxic molecule. Such domain-complementary molecule fusion proteins thus represent a generic targeting vehicle for cell/tissue-specific delivery of generic anti-complementary-detectable/ cytotoxic molecule conjugates.

In another embodiment, znssp6-cytokine fusion proteins or antibody-cytokine fusion proteins can be used for enhancing *in vivo* killing of target tissues (for example, brain, kidney, testis, and other tissues), if the znssp6 polypeptide or anti-znssp6 antibody targets, for example, the hyperproliferative brain, kidney, testis (See, generally, Hornick et al., Blood 89:4437-47, 1997). They described fusion proteins

which enable targeting of a cytokine to a desired site of action, thereby providing an elevated local concentration of cytokine. Suitable znssp6 polypeptides or anti-znssp6 antibodies target an undesirable cell or tissue (i.e., a tumor or a leukemia), and the fused cytokine mediated improved target cell lysis by effector cells. Suitable cytokines for this purpose include interleukin 2 and granulocyte-macrophage colony-stimulating factor (GM-CSF), for instance.

The bioactive polypeptide or antibody conjugates described herein can be delivered intravenously, intraarterially or intraductally, or may be introduced locally at the intended site of action.

znssp6 polynucleotides and/or polypeptides may be useful for regulating the maturation of UDP-glycosyltransferase substrate-bearing cells, such as fibroblasts, lymphocytes and hematopoietic cells. znssp6 polypeptides will also find use in mediating metabolic or physiological processes *in vivo*. The effects of a compound on proliferation and differentiation can be measured *in vitro* using cultured cells. Bioassays and ELISAs are available to measure cellular response to znssp6, in particular are those which measure changes in cytokine production as a measure of cellular response (see for example, Current Protocols in Immunology ed. John E. Coligan et al., NIH, 1996). Assays to measure other cellular responses, including glycoprotein and glycolipid biosynthesis and metabolism, and cell-cell interactions are known in the art.

The activity of znssp6 or a peptide to which znssp6 binds, can be measured by a silicon-based biosensor microphysiometer which measures the extracellular acidification rate or proton excretion associated with such protein interactions and subsequent physiologic cellular responses. An exemplary device is the Cytosensor™ Microphysiometer manufactured by Molecular Devices, Sunnyvale, CA. A variety of cellular responses, such as cell proliferation, ion transport, energy production, inflammatory response, regulatory and enzyme or enzyme activation, and the like, can be measured by this method. See, for example, McConnell, H.M. et al., Science 257:1906-1912, 1992; Pitchford, S. et al., Meth. Enzymol. 228:84-108, 1997; Arimilli, S. et al., J. Immunol. Meth. 212:49-59, 1998; Van Liefde, I. et al., Eur. J. Pharmacol. 346:87-95, 1998. The microphysiometer can be used for assaying adherent

or non-adherent eukaryotic or prokaryotic cells. By measuring extracellular acidification changes in cell media over time, the microphysiometer directly measures cellular responses to various stimuli, including znssp6 proteins, their agonists, and antagonists. The microphysiometer can be used to measure responses of a znssp6-responsive eukaryotic cell, compared to a control eukaryotic cell that does not respond to znssp6 polypeptide. znssp6-responsive eukaryotic cells comprise cells into which a polynucleotide for znssp6 has been transfected creating a cell that is responsive to znssp6; or cells containing endogenous znssp6 polynucleotides. Differences, measured by a change in the response of cells exposed to znssp6 anti-complementary molecule, relative to a control not exposed to znssp6 anti-complementary molecule, directly measure the znssp6-modulated cellular responses. Moreover, such znssp6-modulated responses can be assayed under a variety of stimuli. The present invention provides a method of identifying agonists and antagonists of znssp6 protein, comprising providing cells responsive to a znssp6 polypeptide, culturing a first portion of the cells in the absence of a test compound, culturing a second portion of the cells in the presence of a test compound, and detecting a measurable change in a cellular response of the second portion of the cells as compared to the first portion of the cells. Moreover, culturing a third portion of the cells in the presence of znssp6 anti-complementary molecule and the absence of a test compound provides a positive control for the znssp6-responsive cells, and a control to compare the agonist activity of a test compound with that of the znssp6 anti-complementary molecule. Antagonists of znssp6 can be identified by exposing the cells to znssp6 anti-complementary molecule in the presence and absence of the test compound, whereby a reduction in znssp6-modulated activity is indicative of antagonist activity in the test compound.

Moreover, znssp6 can be used to identify cells, tissues, or cell lines which respond to a znssp6-modulated pathway. The microphysiometer, described above, can be used to rapidly identify cells responsive to znssp6 of the present invention. Cells can be cultured in the presence or absence of znssp6 polypeptide. Those cells which elicit a measurable change in extracellular acidification in the presence of znssp6 are responsive to znssp6. Such cell lines, can be used to identify

znssp6 anti-complementary molecule, antagonists and agonists of znssp6 polypeptide as described above.

Molecules of the present invention can be used to identify and isolate receptors, ligands, or members of complement/anti-complement pairs involved in cell-cell interactions, and glycoprotein synthesis. For example, proteins and peptides of the present invention can be immobilized on a column and membrane preparations run over the column (Immobilized Affinity Ligand Techniques, Hermanson et al., eds., Academic Press, San Diego, CA, 1992, pp.195-202). Proteins and peptides can also be radiolabeled (Methods in Enzymol., vol. 182, "Guide to Protein Purification", M. Deutscher, ed., Acad. Press, San Diego, 1990, 721-37) or photoaffinity labeled (Brunner et al., Ann. Rev. Biochem. 62:483-514, 1993 and Fedan et al., Biochem. Pharmacol. 33:1167-80, 1984) and specific cell-surface proteins can be identified.

As a reagent, the polynucleotide encoding the amino acid residues from residue 304 to 309 of SEQ ID NO: 2, and the degenerate polynucleotide of SEQ ID NO:3, can be used to identify new family members. This would be useful in finding new galactosyltransferase and putative neurogenic secreted signaling peptides from the same or other tissues.

The polypeptides, nucleic acid and/or antibodies of the present invention can be used in treatment of disorders associated with cell migration, contact inhibition, tissue interactions, neuronal specificity, fertilization, embryonic cell adhesions, limb bud morphogenesis, mesenchyme development, immune recognition, growth control, tumor metastasis, and glycoprotein and glycolipid biosynthesis. The molecules of the present invention can be used to modulate glycoprotein synthesis and/or cell-cell interactions or to treat or prevent development of pathological conditions in such diverse tissue as brain, kidney, testis, spinal cord, colon, prostate, stomach, ovary, pancreas, pituitary gland, adrenal gland, salivary gland, mammary gland, liver, small intestine, spleen, thymus, peripheral leukocyte, lymph node, bone marrow, lung, trachea, placenta, fetal spleen and fetal lung. In particular, certain neuronal and epithelial deficiencies and malignancies may be amenable to such diagnosis, treatment or prevention.

Polynucleotides encoding znssp6 polypeptides are useful within gene therapy applications where it is desired to increase or inhibit znssp6 activity. If a mammal has a mutated or absent znssp6 gene, the znssp6 gene can be introduced into the cells of the mammal. In one embodiment, a gene encoding a znssp6 polypeptide is introduced *in vivo* in a viral vector. Such vectors include an attenuated or defective DNA virus, such as, but not limited to, herpes simplex virus (HSV), papillomavirus, Epstein Barr virus (EBV), adenovirus, adeno-associated virus (AAV), and the like. Defective viruses, which entirely or almost entirely lack viral genes, are preferred. A defective virus is not infective after introduction into a cell. Use of defective viral vectors allows for administration to cells in a specific, localized area, without concern that the vector can infect other cells. Examples of particular vectors include, but are not limited to, a defective herpes simplex virus 1 (HSV1) vector (Kaplit et al., Molec. Cell. Neurosci. 2:320-30, 1991); an attenuated adenovirus vector, such as the vector described by Stratford-Perricaudet et al., J. Clin. Invest. 90:626-30, 1992; and a defective adeno-associated virus vector (Samulski et al., J. Virol. 61:3096-101, 1987; Samulski et al., J. Virol. 63:3822-8, 1989).

In another embodiment, a znssp6 gene can be introduced in a retroviral vector, e.g., as described in Anderson et al., U.S. Patent No. 5,399,346; Mann et al. Cell 33:153, 1983; Temin et al., U.S. Patent No. 4,650,764; Temin et al., U.S. Patent No. 4,980,289; Markowitz et al., J. Virol. 62:1120, 1988; Temin et al., U.S. Patent No. 5,124,263; International Patent Publication No. WO 95/07358, published March 16, 1995 by Dougherty et al.; and Kuo et al., Blood 82:845, 1993.

Alternatively, the vector can be introduced by lipofection *in vivo* using liposomes. Synthetic cationic lipids can be used to prepare liposomes for *in vivo* transfection of a gene encoding a marker (Felgner et al., Proc. Natl. Acad. Sci. USA 84:7413-7, 1987; Mackey et al., Proc. Natl. Acad. Sci. USA 85:8027-31, 1988). The use of lipofection to introduce exogenous genes into specific organs *in vivo* has certain practical advantages. Molecular targeting of liposomes to specific cells represents one area of benefit. More particularly, directing transfection to particular cells represents one area of benefit. For instance, directing transfection to particular cell types would be particularly advantageous in a tissue with cellular heterogeneity, such as the pancreas,

liver, kidney, and brain. Lipids may be chemically coupled to other molecules for the purpose of targeting. Targeted peptides (e.g., hormones or neurotransmitters), proteins such as antibodies, or non-peptide molecules can be coupled to liposomes chemically. Similarly, the znssp6 polynucleotide itself can be used to target specific tissues.

5 It is possible to remove the target cells from the body; to introduce the vector as a naked DNA plasmid; and then to re-implant the transformed cells into the body. Naked DNA vectors for gene therapy can be introduced into the desired host cells by methods known in the art, e.g., transfection, electroporation, microinjection, transduction, cell fusion, DEAE dextran, calcium phosphate precipitation, use of a gene
10 gun or use of a DNA vector transporter. See, e.g., Wu et al., J. Biol. Chem. 267:963-7, 1992; Wu et al., J. Biol. Chem. 263:14621-4, 1988.

Various techniques, including antisense and ribozyme methodologies, can be used to inhibit znssp6 gene transcription and translation, such as to inhibit cell proliferation in vivo. Polynucleotides that are complementary to a segment of a
15 znssp6-encoding polynucleotide (e.g., a polynucleotide as set forth in SEQ ID NO:1) are designed to bind to znssp6-encoding mRNA and to inhibit translation of such mRNA. Such antisense polynucleotides are used to inhibit expression of znssp6 polypeptide-encoding genes in cell culture or in a subject.

The present invention also provides reagents which will find use in
20 diagnostic applications. For example, the znssp6 gene, a probe comprising znssp6 DNA or RNA or a subsequence thereof can be used to determine if the znssp6 gene is present on chromosome 12q24.31 or if a mutation has occurred. Detectable chromosomal aberrations at the znssp6 gene locus include, but are not limited to, aneuploidy, gene copy number changes, insertions, deletions, restriction site changes
25 and rearrangements. Such aberrations can be detected using polynucleotides of the present invention by employing molecular genetic techniques, such as restriction fragment length polymorphism (RFLP) analysis, short tandem repeat (STR) analysis employing PCR techniques, and other genetic linkage analysis techniques known in the art (Sambrook et al., *ibid.*; Ausubel et al., *ibid.*; Marian, Chest 108:255-65, 1995).

30 In general, these diagnostic methods comprise the steps of (a) obtaining a genetic sample from a patient; (b) incubating the genetic sample with a

polynucleotide probe or primer as disclosed above, under conditions wherein the polynucleotide will hybridize to complementary polynucleotide sequence, to produce a first reaction product; and (iii) comparing the first reaction product to a control reaction product. A difference between the first reaction product and the control reaction product is indicative of a genetic abnormality in the patient. Genetic samples for use within the present invention include genomic DNA, cDNA, and RNA. The polynucleotide probe or primer can be RNA or DNA, and will comprise a portion of SEQ ID NOs:1 or 3, the complement of SEQ ID NOs:1 or 3, or an RNA equivalent thereof. Suitable assay methods in this regard include molecular genetic techniques known to those in the art, such as restriction fragment length polymorphism (RFLP) analysis, short tandem repeat (STR) analysis employing PCR techniques, ligation chain reaction (Barany, PCR Methods and Applications 1:5-16, 1991), ribonuclease protection assays, and other genetic linkage analysis techniques known in the art (Sambrook et al., ibid.; Ausubel et. al., ibid.; Marian, Chest 108:255-65, 1995). Ribonuclease protection assays (see, e.g., Ausubel et al., ibid., ch. 4) comprise the hybridization of an RNA probe to a patient RNA sample, after which the reaction product (RNA-RNA hybrid) is exposed to RNase. Hybridized regions of the RNA are protected from digestion. Within PCR assays, a patient's genetic sample is incubated with a pair of polynucleotide primers, and the region between the primers is amplified and recovered. Changes in size or amount of recovered product are indicative of mutations in the patient. Another PCR-based technique that can be employed is single strand conformational polymorphism (SSCP) analysis (Hayashi, PCR Methods and Applications 1:34-8, 1991).

In addition, such polynucleotide probes could be used to hybridize to counterpart sequences on individual chromosomes. Chromosomal identification and/or mapping of the znssp6 gene could provide useful information about gene function and disease association. Many mapping techniques are available to one skilled in the art, for example, mapping somatic cell hybrids, and fluorescence *in situ* hybridization (FISH). One method is radiation hybrid mapping. Radiation hybrid mapping is a somatic cell genetic technique developed for constructing high-resolution, contiguous maps of mammalian chromosomes (Cox et al., Science 250:245-50, 1990). Partial or

full knowledge of a gene's sequence allows one to design PCR primers suitable for use with chromosomal radiation hybrid mapping panels. Radiation hybrid mapping panels are commercially available which cover the entire human genome, such as the Stanford G3 RH Panel and the GeneBridge 4 RH Panel (Research Genetics, Inc., Huntsville, AL). These panels enable rapid, PCR-based chromosomal localizations and ordering of genes, sequence-tagged sites (STSs), and other nonpolymorphic and polymorphic markers within a region of interest. This includes establishing directly proportional physical distances between newly discovered genes of interest and previously mapped markers. The precise knowledge of a gene's position can be useful for a number of purposes, including: 1) determining if a sequence is part of an existing contig and obtaining additional surrounding genetic sequences in various forms, such as YACs, BACs or cDNA clones; 2) providing a possible candidate gene for an inheritable disease which shows linkage to the same chromosomal region; and 3) cross-referencing model organisms, such as mouse, which may aid in determining what function a particular gene might have.

Sequence tagged sites (STSs) can also be used independently for chromosomal localization. An STS is a DNA sequence that is unique in the human genome and can be used as a reference point for a particular chromosome or region of a chromosome. An STS is defined by a pair of oligonucleotide primers that are used in a polymerase chain reaction to specifically detect this site in the presence of all other genomic sequences. Since STSs are based solely on DNA sequence they can be completely described within an electronic database, for example, Database of Sequence Tagged Sites (dbSTS), GenBank, (National Center for Biological Information, National Institutes of Health, Bethesda, MD <http://www.ncbi.nlm.nih.gov>), and can be searched with a gene sequence of interest for the mapping data contained within these short genomic landmark STS sequences.

Transgenic mice, engineered to express the znssp6 gene, and mice that exhibit a complete absence of znssp6 gene function, referred to as "knockout mice" (Snouwaert et al., *Science* 257:1083, 1992), may also be generated (Lowell et al., *Nature* 366:740-42, 1993). These mice may be employed to study the znssp6 gene,

cells affected by the znssp6 gene product, and the protein encoded thereby in an *in vivo* system.

Znssp6 polypeptides, variants, and fragments thereof, may be useful as replacement therapy for disorders associated with glycoprotein synthesis and cell-cell interactions.

Brn is thought to be a signaling molecule for a novel receptor, thus regulating adhesion between germ and follicle cells. It is also theorized that *brn* may be required for lateral inhibition during early neurogenesis, maintaining epithelial structure within the neurogenic ectoderm during neuroblast segregation, and is necessary for epithelial maintenance. See Goode, S. et al., Dev. Biol. 178:35-50, 1996. Since the *Drosophila* follicular epithelium has developmental, morphological, and molecular properties of vertebrate epithelia, (Goode, S. et al., Development 122:3863-3879, 1996) it is considered likely that its human orthologue would be involved in epithelial development.

The protein of the present invention has 36% identity to *brn*. High expression of znssp6 is observed in fetal and adult brain. This suggests that znssp6 plays a role in epithelial and neuronal development and maintenance. It's role in embryonic development as well as differentiation and stability throughout life may be critical.

Additionally, it has been proposed (Goode, et al., 1996, *ibid*) that *brn* and another neurogenic secreted signaling peptide, *Egghead*, collaborate with yet a third neurogenic secreted signaling peptide, *Notch*, on the apical surface of follicle cells to mediate germline follicle cell adhesion. Thus, znssp6 may act on cell-cell contact alone, or in collaboration with other surface molecules.

Mutants of these neurogenic secreted signaling peptides, *brn*, *Egghead*, and *Notch*, result in follicle cells which over proliferate (Goode, S. et al., *ibid*). Thus, these peptides, and the genes encoding them can play a role in tumor suppression. It is likely that homologous proteins, such as znssp6, can mediate tumor suppression as well.

The protein of the present invention has 43 % homology to Cardiac and Pancreatic Protein (WO 98/44112) which is contemplated to exert an effect on the

differentiation of cells in early stages of cell and tissue development, and possibly serves to aid in the differentiation of embryonic cells into heart and pancreas cells. Similarly, znssp6 may affect the differentiation of cells in the early stages of cell and tissue development. Znssp6 may also affect the differentiation of embryonic cells into
 5 brain, kidney, and testis cells, or the cells and tissues such as spinal cord, colon, prostate, stomach, ovary, pancreas, pituitary gland, adrenal gland, salivary gland, mammary gland, liver, small intestine, spleen, thymus, peripheral leukocyte, lymph node, bone marrow, lung, trachea, placenta, fetal spleen and fetal lung.

A less widely appreciated determinant of tissue morphogenesis is the
 10 process of cell rearrangement: Both cell motility and cell-cell adhesion are likely to play central roles in morphogenetic cell rearrangements. Cells need to be able to rapidly break and probably simultaneously remake contacts with neighboring cells. See Gumbiner, B.M., Cell 69:385-387, 1992. As a secreted protein in brain, spinal cord, kidney, and testes, znssp6 can play a role in intercellular rearrangement in these and
 15 other tissues.

The znssp6 polypeptide is expressed in tissues of the brain, kidney and testis. Thus, the polypeptides of the present invention are useful in studying cell adhesion and the role thereof in metastasis and may be useful in preventing metastasis, in particular metastasis in tumors of the brain, kidney and testis. Similarly,
 20 polynucleotides and polypeptides of znssp6 may be used to replace their defective counterparts in tumor or diseased tissues. Thus, znssp6 polypeptide pharmaceutical compositions of the present invention may be useful in prevention or treatment of disorders associated with pathological regulation or the expansion of these tissues. The polynucleotides of the present invention may also be used in conjunction with a
 25 regulatable promoter, thus allowing the dosage of delivered protein to be regulated.

Moreover, the activity and effect of znssp6 on tumor progression and metastasis can be measured *in vivo*. Several syngeneic mouse models have been developed to study the influence of polypeptides, compounds or other treatments on tumor progression. In these models, tumor cells passaged in culture are implanted into
 30 mice of the same strain as the tumor donor. The cells will develop into tumors having similar characteristics in the recipient mice, and metastasis will also occur in some of

the models. Tumor models include the Lewis lung carcinoma (ATCC No. CRL-1642) and B16 melanoma (ATCC No. CRL-6323), amongst others. These are both commonly used tumor lines, syngeneic to the C57BL6 mouse, that are readily cultured and manipulated *in vitro*. Tumors resulting from implantation of either of these cell lines are capable of metastasis to the lung in C57BL6 mice. The Lewis lung carcinoma model has recently been used in mice to identify an inhibitor of angiogenesis (O'Reilly MS, et al. Cell 79: 315-328,1994). C57BL6/J mice are treated with an experimental agent either through daily injection of recombinant protein, agonist or antagonist or a one time injection of recombinant adenovirus. Three days following this treatment, 10^5 to 10^6 cells are implanted under the dorsal skin. Alternatively, the cells themselves may be infected with recombinant adenovirus, such as one expressing znssp6, before implantation so that the protein is synthesized at the tumor site or intracellularly, rather than systemically. The mice normally develop visible tumors within 5 days. The tumors are allowed to grow for a period of up to 3 weeks, during which time they may reach a size of 1500 - 1800 mm³ in the control treated group. Tumor size and body weight are carefully monitored throughout the experiment. At the time of sacrifice, the tumor is removed and weighed along with the lungs and the liver. The lung weight has been shown to correlate well with metastatic tumor burden. As an additional measure, lung surface metastases are counted. The resected tumor, lungs and liver are prepared for histopathological examination, immunohistochemistry, and *in situ* hybridization, using methods known in the art and described herein. The influence of the expressed polypeptide in question, e.g., znssp6, on the ability of the tumor to recruit vasculature and undergo metastasis can thus be assessed. In addition, aside from using adenovirus, the implanted cells can be transiently transfected with znssp6. Moreover, purified znssp6 or znssp6-conditioned media can be directly injected in to this mouse model, and hence be used in this system. Use of stable znssp6 transfectants as well as use of inducible promoters to activate znssp6 expression *in vivo* are known in the art and can be used in this system to assess znssp6 induction of metastasis. For general reference see, O'Reilly MS, et al. Cell 79:315-328, 1994; and Rusciano D, et al. Murine Models of Liver Metastasis. Invasion Metastasis 14:349-361, 1995.

Tn-syndrome, also called Permanent Mixed-Field Polyagglutinability, is a very rare acquired disorder affecting all hematopoietic lineages. This syndrome is characterized by the expression of the Tn and sialosyl-Tn antigens on the cell surface. The Tn antigen has been identified as an unsubstituted α -linked *N*-acetyl-galactosamine linked *O*-glycosidically to threonine or serine residues of membrane proteins. In healthy blood, this sugar is substituted by galactose and sialic acid to form a tetrasaccharide. This Tn antigen may be a result of a deficiency in beta-1,3-galactosyltransferase. Expression of the Tn antigen along with the sialosyl-Tn antigen and a TF antigen (characterized by a deficiency in alpha-2,3-sialyltransferase) have been recognized as a cancer-associated phenomenon for many years. See Berger, E.G. et al., Transfus. Clin. Biol. 2:103-108, 1994.

Thus, the study of this syndrome has been useful in elucidating the biology of carbohydrate glycosylation disorders and the appearance of cryptantigens on the cell surface, and cancer. Highly specific and complex tumor glycan antigens are likely of great interest in studying tissue specific tumors and znssp6 can be useful for studying tumors in brain, kidney, and testis tissues.

The expression of these cryptantigens in tissues from normal, chronic pancreatitic, and pancreatic cancer patients was studied by Itzkowitz, et al, Gastroenterology 100:1691-1700, 1991 . The sialosyl-Tn antigen is expressed in 97% of malignant, but 0% of normal tissues. The authors suggest that normal pancreas tissue is preferentially galactosylated resulting in less sialosyl-Tn antigen. In malignant tissue, conditions favor the sialylation of Tn antigens thereby accounting for enhanced expression of sialosyl Tn over T antigens. A similar sialylation of tissues in the brain, kidney, and testis could be associated with disease in these tissues.

In view of the high expression of znssp6 in the in normal tissues of the brain, kidney, and testis, a defect in the znssp6 gene may result in defective galactosylation of cell surface carbohydrates of brain, kidney and testis tissues, leading to over sialylation of the Tn antigen. Thus, znssp6 polypeptides would be useful as a brain, kidney or testis beta-1,3-galactosyltransferase replacement therapy for pre-cancerous and cancer tissues. To verify the presence of such activity in znssp6 containing normal cell lines and tumor cell lines, such cell lines are evaluated with

respect to the presence of the Tn antigen according to procedures known in the art. See, for example, Berger et al., *ibid.*, Itzkowitz et al., *ibid.* and the like.

Additionally, the lack of conditions favoring proper galactosylation may result in an increase in sialosyl Tn antigens in tissues expressing znssp6, which may
5 cause an auto-immune reaction resulting in an immune attack on the brain, kidney, and testis. In these cases, znssp6 molecules may be used to encourage proper galactosylation and limit the antigenic recognition in tissues over expressing the sialosyl Tn antigen.

Similarly, a defective znssp6 gene may result in improper glycoslation
10 of the surface carbohydrates of the tissues of brain, kidney, and testis, thus affecting cell-cell interactions and possibly cell cycle regulation. Such cases could be treated by administering polypeptides of znssp6 to mammals with such a defective gene.

Znssp6 gene may be useful as a probe to identify humans who have a defective znssp6 gene. The strong expression of znssp6 in brain, kidney, and testis
15 suggests that znssp6 polynucleotides or polypeptides are down-regulated in tumor or malignant tissues. Thus, polynucleotides and polypeptides of znssp6, and mutations to them, can be used a diagnostic indicators of cancer in these tissues. Thus, polynucleotides and polypeptides of znssp6, and mutations to them, can be used a indicators of pancreatic and colonic cancer, and disease, in diagnosis.

The polypeptides, nucleic acid, and/or antibodies of the present
20 invention may be used in treatment of disorders associated with brain, kidney, and testis activity and in disorders associated with glycoprotein synthesis. The molecules of the present invention may used to modulate or to treat or prevent development of pathological conditions in such diverse tissue as brain, kidney, testis, spinal cord,
25 colon, prostate, stomach, ovary, pancreas, pituitary gland, adrenal gland, salivary gland, mammary gland, liver, small intestine, spleen, thymus, peripheral leukocyte, lymph node, bone marrow, lung, trachea, placenta, fetal spleen and fetal lung. In particular, certain syndromes or diseases may be amenable to such diagnosis, treatment or prevention.

The znssp6 polypeptide is expressed in the brain, kidney, and testis.
30 Thus, znssp6 polypeptide pharmaceutical compositions of the present invention may be

useful in prevention or treatment of disorders associated with pathological regulation or the expansion of brain, kidney, and testis tissues.

The polynucleotides of the present invention may also be used in conjunction with a regulatable promoter, thus allowing the dosage of delivered protein to be regulated.

The localization of znssp6 to chromosome 12q24 maps this gene to the same region as a disease known as Adult Spinal Muscular Atrophy (also known as Spinal Muscular Atrophy IV (SMA IV); Hereditary Motor Neuropathy, Distal Included (HMN, included); and HMN, Distal, Type II, Included (HMN2, included)). This disease is characterized by rapid progression of a form of Spinal Muscular Atrophy beginning between the end of the fourth and sixth decades. More information on this disease can be found on the internet on the Online Mendelian Inheritance of Man homepage (<http://www3.ncbi.nlm.nih.gov/Omim/>). Thus, znssp6 could be the gene causing Adult Spinal Muscular Atrophy. In such case, therapy with znssp6 as replacement of the polypeptide or polynucleotide would be useful.

For pharmaceutical use, the proteins of the present invention can be administered orally, rectally, parenterally, intracisternally, intravaginally, intraperitoneally, topically (as powders, ointments, drops or transdermal patch) buccally, or as a pulmonary or nasal inhalant. Intravenous administration will be by bolus injection or infusion over a typical period of one to several hours. In general, pharmaceutical formulations will include a znssp6 protein in combination with a pharmaceutically acceptable vehicle, such as saline, buffered saline, 5% dextrose in water or the like. Formulations may further include one or more excipients, preservatives, solubilizers, buffering agents, albumin to prevent protein loss on vial surfaces, etc. Methods of formulation are well known in the art and are disclosed, for example, in Remington: The Science and Practice of Pharmacy, Gennaro, ed., Mack Publishing Co., Easton, PA, 19th ed., 1995. Therapeutic doses will generally be in the range of 0.1 to 100 µg/kg of patient weight per day, preferably 0.5-20 mg/kg per day, with the exact dose determined by the clinician according to accepted standards, taking into account the nature and severity of the condition to be treated, patient traits, etc. Determination of dose is within the level of ordinary skill in the art. The proteins may

be administered for acute treatment, over one week or less, often over a period of one to three days or may be used in chronic treatment, over several months or years. In general, a therapeutically effective amount of znssp6 is an amount sufficient to produce a clinically significant change in brain, kidney and testis tissues. Similarly, a therapeutically effective amount of znssp6 is an amount sufficient to produce a clinically significant change in disorders associated with spinal cord, colon, prostate, stomach, ovary, pancreas, pituitary gland, adrenal gland, salivary gland, mammary gland, liver, small intestine, spleen, thymus, peripheral leukocyte, lymph node, bone marrow, lung, trachea, placenta, fetal spleen and fetal lung.

The invention is further illustrated by the following non-limiting examples.

EXAMPLES

Example 1

Extension of EST Sequence

The polynucleotide sequence of the novel polypeptides of the present invention was initially identified by querying an EST database. A cDNA clone, corresponding to an EST was obtained and the deduced amino acid sequence of the insert was determined to be incomplete at the 5' terminal. Nested 5' RACE polymerase chain reactions were performed using human kidney marathon cDNAs. The first RACE used primers ZC9739(SEQ ID NO:4) and ZC17164 (SEQ ID NO:5) and thermalcycler conditions as follows: one cycle of 94°C for 2 minutes; followed by thirty cycles of 94°C for 20 seconds, 72°C for 1 minute; followed by one cycle of 72°C for 2 minutes. The second, nested, RACE reaction used diluted product from the first reaction and primers ZC9719 (SEQ ID NO:6) and ZC17165 (SEQ ID NO:7) and thermalcycler conditions as follows: one cycle of 94°C for 2 minutes; followed by five cycles of 94°C for 20 seconds, 66°C for 30 seconds, 72°C for 1 minute; followed by twenty-five cycles of 94°C for 20 seconds, 64°C for 30 seconds, 72°C for 1 minute; followed by one cycle at 72°C for 7 minutes. Comparison of the 5' extension of the EST sequence with other family members indicated that the 5' RACE product was still not full length. A REX analysis performed on the newly generated 5' sequence data,

identified a second EST which overlaps with the 5' RACE sequence. The insert corresponding to this second EST was determined to be full-length znssp6.

One skilled in the art would be able to isolate the full-length polynucleotide in the following manner: Sense and antisense oligonucleotides can be designed to encompass the 5' and 3' ends of the polynucleotide sequence, respectively. Exemplary oligonucleotides would be a sense primer (SEQ. ID NO:8), and an antisense primer (SEQ. ID NO:9). cDNA from a kidney library can be used as template, and the following thermalcycler conditions can be used: 94 degrees for 2 minutes; followed by thirty cycles of 94 degrees for 20 seconds, 72 degrees for 1 minute; followed by a final extension of 72 degrees for 7 minutes. The resulting PCR product can be subcloned and sequenced.

Alternatively, one can order the clone corresponding to the second EST, number 188406, from IMAGE Consortium (info@image.llnl.gov).

Example 2

15 Tissue Distribution

Analysis of tissue distribution was performed by the Northern blotting technique using Human Multiple Tissue and Master Dot Blots (Clontech, Palo Alto, CA). A probe of about 430 base pairs was obtained by PCR of the original EST template using primers ZC17161 (SEQ ID NO:10) and ZC17160 (SEQ ID NO:11). Thermalcycler conditions were as follows: one cycle of 94°C for 2 minutes; followed by thirty-five cycles of 94°C for 20 seconds, 65°C for 30 seconds, 72°C for 30 seconds; followed by one cycle of 72°C for 2 minutes. The PCR product was gel-purified and random prime labeled with ³²P using a commercially available kit (Rediprime DNA Labeling System; Amersham Corp., Arlington Heights, IL) according to the manufacturer's direction. The probe was then purified using a NucTrap® probe purification column (Stratagene, La Jolla, CA). ExpressHyb™ Hybridization Solution (Clontech, Palo Alto, CA) was used for pre-hybridization and hybridization. Hybridization took place overnight at 65°C, and the blots were then washed three times in 2X SSC and 0.05% SDS at 55°C, followed by two washes in 0.1X SSC and 0.1% SDS at 55°C. The blots were then exposed to autoradiograph film, which was then

developed. Strong signals were observed in brain (adult and fetal), kidney (adult and fetal), and testis. The major transcript size was about 1.6 kb while minor transcripts of about 3.0 kb, and 4.0 kb were also evident. Lower level expression was also observed in tissues such as spinal cord, colon, prostate, stomach, ovary, pancreas, pituitary gland, adrenal gland, salivary gland, mammary gland, liver, small intestine, spleen, thymus, peripheral leukocyte, lymph node, bone marrow, lung, trachea, placenta, fetal spleen and fetal lung.

Example 3

Chromosomal Assignment and Placement of Znssp6

Znssp6 was mapped to chromosome 12 using the commercially available version of the "Stanford G3 Radiation Hybrid Mapping Panel" (Research Genetics, Inc., Huntsville, AL). The "Stanford G3 RH Panel" contains DNA from each of 83 radiation hybrid clones of the whole human genome, plus two control DNAs (the RM donor and the A3 recipient). A publicly available WWW server (<http://shgc-www.stanford.edu>) allows chromosomal localization of markers.

For the mapping of znssp6 with the "Stanford G3 RH Panel", 20 μ l reactions were set up in a 96-well microtiter plate compatible for PCR (Stratagene, La Jolla, CA) and used in a "RoboCycler Gradient 96" thermal cycler (Stratagene). Each of the 85 PCR reactions consisted of 2 μ l 10X KlenTaq PCR reaction buffer (CLONTECH Laboratories, Inc., Palo Alto, CA), 1.6 μ l dNTPs mix (2.5 mM each, PERKIN-ELMER, Foster City, CA), 1 μ l sense primer, ZC 19,220 (SEQ ID NO:12), 1 μ l antisense primer, ZC 19,221 (SEQ ID NO:13), 2 μ l "RediLoad" (Research Genetics, Inc., Huntsville, AL), 0.4 μ l 50X Advantage KlenTaq Polymerase Mix (Clontech Laboratories, Inc.), 25 ng of DNA from an individual hybrid clone or control and distilled water for a total volume of 20 μ l. The reactions were overlaid with an equal amount of mineral oil and sealed. The PCR cycler conditions were as follows: one cycle of 94°C for 5 minutes; followed by 35 cycles of 94 for 45 seconds, 68°C for 45 seconds, and 72°C for 1 minute, 15 seconds; followed by a final extension of 7 minutes at 72°C. The reactions were separated by electrophoresis on a 2% agarose gel (Life Technologies, Gaithersburg, MD).

The results showed linkage of znssp6 to the framework marker SHGC-13898 with a LOD score of >9 and at a distance of 19.43 cR_10000 from the marker. The use of surrounding genes and/or markers positions znssp6 in the 12q24.

5 Example 4

Construction of znssp6 Glu-Glu-Tagged Expression Vectors for *Pichia methanolica*

Expression of znssp6 in *Pichia methanolica* utilizes the expression system described in co-assigned WIPO publication WO 97/17450. An expression plasmid containing all or part of a polynucleotide encoding znssp6 is constructed via
 10 homologous recombination. The expression vector is built from pCZR190, which contains the AUG1 promoter, followed by the alpha factor prepro (α Fpp) leader sequence, followed by an amino-terminal Glu-Glu tag, a blunt-ended Sma I restriction site for insertion of the gene sequence of interest, a translational stop codon, followed by the AUG1 terminator, the ADE2 selectable marker, and finally the AUG1 3'
 15 untranslated region. Also included in this vector are the URA3 and CEN-ARS sequences required for selection and replication in *S. cerevisiae*, and the AmpR and colE1 ori sequences required for selection and replication in *E. coli*. The znssp6 sequence inserted into this vector begins at residue 50 (Arg) of the znssp6 amino acid sequence (SEQ ID NO:2).

20 Gene expression constructs are prepared by PCR and homologously recombined into the yeast expression vector pCZR190. For the amino terminal tagged protein, the N-terminal primer, ZC25565 (SEQ ID NO:14), comprises 40 base pairs containing the α Fpp coding sequence joined to a nucleotide sequence encoding a Glu-Glu tag followed by 25 base pairs of nucleotide sequence encoding a portion of the
 25 amino-terminus from the ectodomain of the znssp6 sequence. The C-terminal primer, ZC25561 (SEQ ID NO:15), comprises about 25 base pairs of carboxy terminus coding sequence of the znssp6 joined with 40 base pairs of AUG1 terminator sequence. The polymerase chain reaction contains 1 ng znssp6 template (i.e., from SEQ ID NO:1), 100 pmol of each primer, 10 μ l of 10X PCR buffer, 1 μ l Pwo polymerase (Boehringer
 30 Mannheim, Indianapolis, IN), 10 μ l of 0.25 mM nucleotide triphosphate mix (Perkin

Elmer, Foster City, CA) and dH₂O to a total volume of 100 μ l. PCR conditions are as follows: 25 cycles of 30 seconds at 94°C, 1 minute at 55°C and 1 minute at 72°C°.

Similarly, for the carboxyl terminal tagged protein, the N-terminal primer, ZC25563 (SEQ ID NO:16), comprises 40 base pairs containing the α Fpp coding sequence joined to 25 base pairs of nucleotide sequence encoding a portion of the amino-terminus from the ectodomain of the znssp6 sequence. The C-terminal primer, ZC25564 (SEQ ID NO:17), comprises about 25 base pairs of carboxy terminus coding sequence of the znssp6 joined to a nucleotide sequence encoding a Glu-Glu tag followed by 40 base pairs of AUG1 terminator sequence. Polymerase chain reaction volumes and conditions are the same as those listed for the amino terminal tagged gene construct.

An untagged gene expression construct is also prepared: the N-terminal primer, ZC25562 (SEQ ID NO:18), comprises 40 base pairs containing the α Fpp coding sequence joined to 25 base pairs of nucleotide sequence encoding a portion of the amino-terminus from the ectodomain of the znssp6 sequence. The C-terminal primer, ZC25566 (SEQ ID NO:19), comprises about 25 base pairs of carboxy terminus coding sequence of the znssp6 joined to 40 base pairs of AUG1 terminator sequence. Polymerase chain reaction volumes and conditions are the same as those listed for the amino terminal tagged gene construct.

The NEE-, CEE- and untagged znssp6 plasmids are made by homologously recombining 100 ng of Sma I digested pCRZ204 acceptor vector, and 1 μ g of Eco RI-Xho I znssp6 gene constructs made above into *S. cerevisiae*. One hundred microliters of competent yeast cells (*S. cerevisiae*) are combined with 10 μ l of each of the fragments and transferred to a 0.2 cm electroporation cuvette. The yeast/DNA mixture is electropulsed at 0.75 kV (5 kV/cm), ∞ ohms, 25 μ F. To the cuvette is added 600 μ l of 1.2 M sorbitol and 300 μ l aliquots of the yeast/sorbitol mixture are plated onto two URA D plates and incubated at 30°C.

After about 48 hours the Ura⁺ yeast transformants from a single plate are resuspended in 2.5 ml H₂O and spun briefly to pellet the yeast cells. The cell pellet is resuspended in 1 ml of lysis buffer (2% Triton X-100, 1% SDS, 100 mM NaCl, 10 mM Tris, pH 8.0, 1 mM EDTA). Five hundred microliters of the lysis mixture was added to

an Eppendorf tube containing 300 μ l acid washed glass beads and 200 μ l phenol-chloroform, vortexed for 1 minute intervals two or three times, followed by a 5 minute spin in a Eppendorf centrifuge at maximum speed. Three hundred microliters of the aqueous phase is transferred to a fresh tube and the DNA precipitated with 600 μ l ethanol (EtOH), followed by a centrifugation at 14,000 RPM of 10 minutes at 4°C. The DNA pellet is resuspended in 100 μ l H₂O.

Five microliters of the resuspended DNA prep is used to transform 40 μ l of electrocompetent *E. coli* cells (DH10B, Gibco BRL). The cells are electropulsed at 2.0 kV and 400 ohms. Following electroporation, 1 ml SOC (2% Bacto™ Tryptone (Difco, Detroit, MI), 0.5% yeast extract (Difco), 10 mM NaCl, 2.5 mM KCl, 10 mM MgCl₂, 10 mM MgSO₄, 20 mM glucose) is added and the cells are allowed to recover for 1 hour at 37°C prior to plating 250 μ l aliquots on four LB AMP plates (LB broth (Lennox), 1.8% Bacto™ Agar (Difco), 100 mg/L Ampicillin).

Individual clones harboring the correct expression construct are identified by PCR screening. The primers used to amplify the N-tagged znssp6 clone are ZC25565 (SEQ ID NO:14) and ZC25561(SEQ ID NO:15). The insert sequence of positive clones, identified by a 1071 bp fragment, are verified by sequence analysis. The primers used to amplify the C-tagged znssp6 clone are ZC25563 (SEQ ID NO:16) and ZC25564(SEQ ID NO:17). The insert sequence of positive clones, identified by a 1067 bp fragment, are verified by sequence analysis. The primers used to amplify the untagged znssp6 clone are ZC25562 (SEQ ID NO:18) and ZC25566(SEQ ID NO:19). The insert sequence of positive clones, identified by a 1066 bp fragment, are verified by sequence analysis. Larger scale plasmid DNA is isolated using Qiagen maxi kits (Qiagen, Valencia, CA) and the DNA is digested with Not I to liberate the *Pichia*-znssp6 expression cassette from the vector backbone. The Not I DNA fragment is then transformed into the *Pichia methanolica* expression host, PMAD16. This is done by mixing 100 μ l of prepared competent PMAD16 cells with 10 ng of Not I digested tagged or untagged znssp6 fragment and transferred to a 0.2 cm electroporation cuvette. The yeast/DNA mixture is electropulsed at 0.75 kV, 25 μ F, infinite ohms. To the cuvette is added 1 ml of 1X Yeast Nitrogen Base and 500 ml aliquots are plated onto two ADE DS (0.056% -Ade -Trp -Thr powder, 0.67% yeast nitrogen base without

amino acids, 2% D-glucose, 0.5% 200X tryptophan, threonine solution, and 18.22% D-sorbitol) plates for selection and incubated at 30°C. Transformants are then picked and screened via Western blot for high-level expression and subjected to large scale fermentation.

- 5 From the foregoing, it will be appreciated that, although specific embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the invention is not limited except as by the appended claims.